# Four New Species of *Amanita* in Inje County, Korea

# Hae Jin Cho<sup>1</sup>, Myung Soo Park<sup>1</sup>, Hyun Lee<sup>1</sup>, Seung-Yoon Oh<sup>1</sup>, Yeongseon Jang<sup>2</sup>, Jonathan J. Fong<sup>3</sup> and Young Woon Lim<sup>1,\*</sup>

<sup>1</sup>School of Biological Sciences, Seoul National University, Seoul 08826, Korea <sup>2</sup>Division of Wood Chemistry & Microbiology, Korea Forest Research Institute, Seoul 02455, Korea <sup>3</sup>Science Unit, Lingnan University, Tuen Mun, New Territories, Hong Kong

**Abstract** Amanita (Agaricales, Basidiomycota) is one of the most well-known genera composed of poisonous mushrooms. This genus of almost 500 species is distributed worldwide. Approximately 240 macrofungi were collected through an ongoing survey of indigenous fungi of Mt. Jeombong in Inje County, Korea in 2014. Among these specimens, 25 were identified as members of *Amanita* using macroscopic features. Specimens were identified to the species level by microscopic features and molecular sequence analyses of the internal transcribed spacer and large subunit of nuclear ribosomal RNA. We molecularly identified 13 *Amanita* species, with seven species matching previously recorded species, four species (A. caesareoides, A. griseoturcosa, A. imazekii, and A. sepiacea) new to Korea, and two unknown species.

Keywords Amanita, Molecular sequence analyses, Mt. Jeombong, New species identification, Poisonous mushrooms

The well-known genus *Amanita* (Agaricales, Basidiomycota) contains both poisonous and edible mushrooms [1]. Species in this genus play important roles in forest ecosystems, as a large majority have a mutualistic association with plants to help effective nutrient uptake [2]. Approximate 500 *Amanita* species have been reported worldwide and are found in a broad range of habitats [3]. *Amanita muscaria* (L.) Lam. is the type species of the genus [4]. Based on morphological features and chemical reaction characters of the fruit body, *Amanita* is divided into two subgenera, including seven sections: *Amanita* Pers. and *Lepidella* (J. E. Gilbert) Veselý emend. Corner & Bas [5]. While this infrageneric classification is not well supported, monophyly of each section, except *Lepidella*, is well supported [5-7].

Nearly 60 Amanita species have been described in Korea

Mycobiology 2015 December, **43**(4): 408-414 http://dx.doi.org/10.5941/MYCO.2015.43.4.408 pISSN 1229-8093 • eISSN 2092-9323 © The Korean Society of Mycology

\*Corresponding author E-mail: ywlim@snu.ac.kr

ReceivedOctober 2, 2015RevisedOctober 12, 2015AcceptedNovember 1, 2015

©This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http:// creativecommons.org/licenses/by-nc/3.0/) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

[8], with most species being reported based on morphological identifications. However, such identification methods are not reliable, and some poisonous *Amanita* species are often confused for edible species [9]. In Korea, 23 deaths due to eating poisonous mushrooms were reported during 2004-2013 according to the Korea Forest Service (http://www.forest.go.kr/). Accurate identification of *Amanita* species is important to reduce these fatal accidents. Molecular phylogenetics is an important tool to help in species identification [6, 10, 11].

Inje County in the Gangwon Province of South Korea is located in the Baekdudaegan Mountains and is known for its rich biodiversity. A total of 523 higher fungi have been reported in Inje County, including 24 *Amanita* species [12]. In 2014, we surveyed Mt. Jeombong in Inje County and identified 25 specimens as *Amanita* species. Specimens were identified to the species level based on morphological features and molecular sequence analysis of the internal transcribed spacer (ITS) and the partial large subunit of nuclear ribosomal RNA (nLSU). Thirteen species were identified, with four species (*A. caesareoides, A. griseoturcosa, A. imazekii*, and *A. sepiacea*) being new records to Korea and two unknown species. Detailed descriptions are presented for the four newly reported *Amanita* species in Korea.

### **MATERIALS AND METHODS**

**Sampling and characterization of morphological features.** All specimens were collected at Mt. Jeombong in Inje County (Gangwon Province, Korea) in 2014.

Species	Collection No.	Best match (accession No.)				Microscopic feature (µm)	
		ITS	Similarity (%)	nLSU	Similarity (%)	Basidia size	Spore size
A. caesareoides <sup>a,b</sup>	SFC20140912-25	A. caesareoides (KP004948)	100	A. caesarea (AF024443)	99.5	42.8~55.0 × 11.5~-14.0	$8.2 \sim 10.8 \times 6.4 \sim 8.5$
	SFC20140822-15						
A. eijii <sup>b</sup>	SFC20140912-04	A. eijii (FJ441039)	96.6	A. cokeri (HQ539682)	95.6	$57.4 \sim 86.8 \times 12.0 \sim 15.2$	$11.1 \sim 13.4 \times 7.2 \sim 8.5$
	SFC20140912-17						
	SFC20141001-14						
A. fulva	SFC20140822-44	A. fulva (AB015692)	99.7	A. fulva (KF021672)	99.8	45.9~66.7 × 15.3~18.3	9.9~12.0 × 9.6~11.8
	SFC20140730-22						
	SFC20140730-23						
A. griseoturcosa <sup>a,b</sup>	SFC20140822-29	A. griseoturcosa (NR 119390)	98.2	A. cylindrispora (AY325867)	95.5	39.1~51.2 × 10.2~12.9	9.1~11.8 × 5.9~7.5
A. imazekii <sup>a,b</sup>	SFC20140912-30	A. imazekii (AB038764)	100	A. yuaniana (AF024488)	94.6	45.8~64.7 × 9.9~14.1	9.8~13.2 × 9.2~11.4
A. manginiana <sup>b</sup>	SFC20140823-10	A. manginiana (KJ466378)	98.7	A. manginiana (AF024463)	100	-	-
A. pallidorosea <sup>b</sup>	SFC20140822-54	A. pallidorosea (KJ739814)	100	A. pallidorosea (KJ466444)	99.5	32.7~49.4 × 8.3~13.2	$7.3 \sim 9.4 \times 6.2 \sim 7.8$
	SFC20140823-08						
A. rubescens	SFC20140822-42	A. rubescens (KF245919)	99.7	A. rubescens (KF245902)	100	26.8~38.7 × 7.4~10.0	7.2~8.6 × 5.3~6.4
	SFC20140823-07						
	SFC20140912-28						
A. sepiacea <sup>a,b</sup>	SFC20140822-49	A. sepiacea (AY436473)	99.8	A. sepiacea (AY436501)	99.0	32.3~40.6 × 8.8~11.1	6.7~8.0 × 4.9~6.8
	SFC20140730-17						
	SFC20140730-25						
	SFC20140912-15						
A. subjunquillea	SFC20140912-03	A. subjunquillea (KJ466427)	100	A. subjunquillea (KF245904)	100	29.4~39.1 × 7.4~11.2	6.7~8.7 × 5.9~8.6
	SFC20140823-11						
A. volvata <sup>b</sup>	SFC20140912-09	A. volvata (AB015681)	91.6	A. volvata (KF245906)	99.7	$42.0 \sim 54.5 \times 10.2 \sim 12.6$	9.6~11.8 × 5.3~7.3
Amanita sp. 1	SFC20140912-22	Amanita sp. (KJ466417)	85.5	A. ponderosa (EF653958)	94.5	-	-
Amanita sp. 2	SFC20140730-10	A. vaginata (AB015691)	93.2	A. angustilamellata (AF024440)	99.4	-	-

Table 1. Amanita specimens examined in this study

<sup>\*</sup>Newly recorded species to Korea. <sup>b</sup>Newly recorded species to Inje County.

#### **410** Cho et al.

Specimens were dried and deposited in the Seoul National University Fungal Collection (SFC) (Table 1). Specimens identified as *Amanita* were re-examined to determine species based on macro- and microscopic characteristics as described in previous studies [1, 13]. After rehydrating in 3% KOH and staining with Congo red solution, microscopic features such as basidia and spores were observed using a light microscope (Eclipse 80i; Nikon, Tokyo, Japan). The morphological features were characterized in detail with specimens that had confirmed identity based on DNA sequence analyses (described below).

# DNA extraction, PCR amplification, and sequencing.

Genomic DNA was extracted from a small piece of tissue using a modified cetyltrimethyl ammonium bromide extraction protocol [14]. The ITS region was amplified with ITS1F and ITS4B primers [15], and the nLSU region was amplified with LR0R (http://www.biology.duke.edu/fungi/mycolab/ primers.htm) and LR5 primers [16]. PCR amplifications were performed on a thermal cycler (C1000TM; Bio-Rad, Richmond, CA, USA) using the AccuPower PCR Premix (Bioneer Co., Daejeon, Korea) following instructions as outlined in Min *et al.* [17]. PCR products were visualized on a 1% agarose gel and purified using the Expin<sup>TM</sup> PCR Purification Kit (GeneAll Biotechnology, Seoul, Korea). Sequencing was performed at Macrogen (Seoul, Korea) on an automated DNA sequencer (ABI3700; Applied Biosystems, Foster City, CA, USA) using the aforementioned PCR primers.

**Sequence analysis.** DNA sequences were proofread using MEGA ver. 5 [18] and aligned with the *Amanita* sequences from the GenBank using Multiple Alignment using Fast Fourier Transform (MAFFT) [19]. Alignments were also checked by eye, and ambiguous positions were adjusted manually. Maximum likelihood (ML) analyses were conducted for the ITS and nLSU datasets, respectively. ML analyses were performed using RAxML ver. 8 [20] with the GTRCAT model of nucleotide substitution and 1,000 bootstrap replicates. For all analyses, *Limacella glioderma* was selected as an outgroup based on a previous study [5]. Intraspecific dissimilarity was calculated using MEGA.



**Fig. 1.** Phylogenetic trees for *Amanita* species based on maximum likelihood analysis of internal transcribed spacer (ITS) and large subunit of nuclear ribosomal RNA (nLSU) sequences. Bootstrap scores of > 50 are presented at the nodes. The scale bar indicates the number of nucleotide substitutions per site. Thickened branches on the root nodes represent identical results between ITS and nLSU, while dotted lines represent the branches which have incongruent results between the ITS and nLSU. Gray boxes indicate the new species found in Korea.

# **RESULTS AND DISCUSSION**

Approximate 240 specimens were collected from Mt. Jeombong in 2014, with 25 identified as Amanita. Amanita specimens were divided into 13 taxa according to their macro- and microscopic features. Basidia and spore size were measured for each specimen, and collection numbers were assigned to all Amanita specimens examined (Table 1). At the first, on the basis of macro- and microscopic features, six taxa were identified: A. fulva, A. manginiana, A. pallidorosea, A. rubescens, A. sepiacea, and A. subjunquillea. The morphological characteristics of each species were well matched with previous referenced descriptions. Molecular identification using both the ITS and nLSU sequences complemented the morphology-based identification. Sequence similarity and phylogenetic analysis confirmed the identity of six Amanita species, in which there was high ITS (over 99.0%) and nLSU (over 99.0%) sequence similarities (Table 1, Fig. 1). However, seven taxa could not be identified at the species level solely based on morphology due to the paucity of distinct traits. In addition, sequence analysis showed some discrepancy of the best match between the ITS and nLSU sequences. Through combining morphological and molecular approaches, five Amanita taxa were further determined at the species level: A. caesareoides, A. eijii, A. griseoturcosa, A. imazekii, and A. volvata. Explanation for the identification of each of the five species follows.

Specimen SFC20140822-15 formed a monophyletic group with A. caesareoides for the ITS sequence (100% sequence similarity) but with A. caesarea for the nLSU sequence (99.5%). Morphologically, A. caesareoides has subglobose to ellipsoid spores [21], while A. caesarea has ellipsoid (occasionally elongate) spores. Specimen SFC20140822-15 had subglobose to broadly ellipsoid spores matching A. caesareoides. Specimen SFC20140912-04 formed a group with A. eijii for the ITS sequence (96.6%) and A. cokeri for the nLSU sequence (95.6%). Morphologically, A. eijii has white to dirty white basidiocarps that become pinkish to brownish in the center, and A. cokeri has white to ivory basidiocarps [22]. Specimen SFC20140912-04 had pinkish to brownish color at the center of basidiocarps, matching A. eijii. Specimen SFC20140822-29 formed a monophyletic clade with the type specimen of A. griseoturcosa in the ITS phylogenetic tree (98.2%) but A. cylindrispora in the nLSU phylogenetic tree (95.5%). Morphologically, these two species are clearly distinguished by their bulb sizes. A. cylindrispora has a longer bulb (50~70 mm) compared to the bulb size of A. griseoturcosa (38~50 mm) [23]. Our specimen has a shorter bulb, which coincides with A. griseoturcosa. Specimen SFC20140912-30 formed a clade with either A. imazekii (ITS, 100%) or A. incarnatifolia (nLSU, 94.6%), and the specimen formed a clade with A. murrilliana (nLSU, 95.0%) and A. yuaniana (nLSU, 94.6%). A. imazekii has globose to subglobose spores [24], while the other three species have the ellipsoid to elongate spores. Specimen SFC20140912-30 had globose to subglobose

spores, identifying it as *A. imazekii*. In the case of specimen SFC20140912-09, its nLSU sequence was highly similar (99.7%) to the sequence of *A. volvata* II (KA12-1194; GenBank: KF245906) of Korea [25]. The low sequence similarity of ITS (91.6%) to *A. volvata* II (KA12-1194, KF245922) and Japanese *A. volvata* (AB015681) [6] raised doubts about its molecular identification. However, morphological characteristics as well as phylogenetic analysis demonstrated that this specimen is indeed *A. volvata* (Fig. 1).

Two immature *Amanita* specimens (SFC20140912-22 and SFC20140730-10) were not confidently identified because morphological features could not be observed and sequence similarity to known species was low (Table 1).

Recently, taxonomic studies based on DNA sequence analysis have increased confidence in identification [17, 26] and showed that Asian fungal species are different from North American and European fungal species with similar morphology [26]. This trend was also found in *Amanita*, with several new species reported from Korea [25, 27]. Many of the previous *Amanita* studies were based solely on morphology, and thus, the true *Amanita* diversity in Korea is unknown. We incorporated molecular phylogenetics in our study to evaluate the diversity of *Amanita* species in Inje County.

Through our surveys of Mt. Jeombong in Inje County, 13 Amanita species were identified using morphological and molecular data (Table 1). Seven of these species were recorded previously in Korea (A. eijii, A. fulva, A. manginiana, A. pallidorosea, A. rubescens, A. subjunquillea, and A. volvata), and four species are new to Korea (A. caesareoides, A. griseoturcosa, A. imazekii, and A. sepiacea) (Fig. 2). Eight of the 13 species are new to the inventory of Amanita in Inje County, increasing species diversity from 24 to 32 species. Inje County has high vegetation diversity, so we expected more new records and possibly new species from the area. Below, we provide a detailed description of four new Amanita species identified in Korea.

#### Taxonomy.

Amanita caesareoides Lj. N. Vassiljeva, Notul. Syst. Sect. Cryptogam. Inst. Bot. Acad. Sci. U. S. S. R. 6: 199 (1950) (Table 1, Fig. 2A~2C).

Cap 68 mm, orangish red to yellow from center to margin. Lamellae yellow to bright yellow, crowded. Stipe  $127 \times 8 \sim 13$  mm, slightly tapering upward, yellow to bright yellow. The bulb is saccate and lobate, outer surface white, 45 mm long and 25 mm wide. The ring is on the upper part of the stem, yellow, skirt-like, membranous. Basidia 42.8~55.0 × 11.5~14.0 µm, 4-sterigmate. Clamps are present at the bases of basidiole. Spores  $8.2 \sim 10.8 \times 6.4 \sim 8.5$  µm, Q =  $1.12 \sim 1.48$ , subglobose to broadly ellipsoidal, occasionally ellipsoidal.

**Specimens examined:** Korea, Gangwon Province, Inje County, Gachilbong and Mt. Jeombong, 38°2'28.56" N, 128°27'58.87" E, 22 Aug 2014, Young Woon Lim,



**Fig. 2.** Four new *Amanita* species identified in Korea. A~C, *A. caesareoides* (SFC20140912-25); D~G, *A. griseoturcosa* (SFC20140822-29); H~J, *A. imazekii* (SFC20140912-30); K~M, *A. sepiacea* (SFC20140822-49). a, spores; b, basidia; c, lamella edge cells (scale bars: B, F, I, L = 2 cm, C, G, J, M = 10  $\mu$ m).

SFC20140822-15; on the ground of mixed woodlands. 37°58′20.76″ N, 128°25′53.15″ E, 12 Sep 2014, Hae Jin Cho, SFC20140912-25 (GenBank accession Nos. KT779079 for ITS and KT779066 for nLSU); on the ground of mixed woodlands.

**Remarks:** Amanita caesareoides is known as the Asian Caesar's mushroom. This species is morphologically similar to *A. caesarea*, *A. hemibapha*, *A. jacksonii*, *A. javanica*, and *A. subjunquillea*. *A. caesareoides* has subglobose to ellipsoid spores, while *A. caesarea*, *A. hemibapha*, *A. jacksonii*, and

*A. javanica* have ellipsoid to elongate spores [21] Among them *A. caesarea*, *A. caesareoides*, and *A. jacksonii* have orange red caps, but they have different geographical distributions. *A. caesareoides* is described in eastern Russian, southwestern China, Korea, and Japan; *A. jacksonii* is described in eastern Canada, the eastern U.S., and eastern Mexico. Also, *A. caesarea* is described from regions near the Mediterranean (studies in the Amanitaceae: http://www.amanitaceae.org/).

A. subjunquillea is one of the lethal Amanita species that

can be misidentified as *A. caesareoides* due to its similar cap color. *A. caesareoides* has prominently pectinate-striate inward margins from the edge of cap and yellow to bright yellow lamellae with subglobose to ellipsoid spores. *A. subjunquillea* has a fine line at the edge of cap and white to cream lamellae with globose to subglobose spores [9]. *A. caesareoides* and *A. subjunquillea* are clearly separated by ITS and nLSU phylogeny.

*Amanita girseoturcosa* T. Oda, C. Tanaka & Tsuda, Mycoscience 43: 351-355 (2002) (Table 1, Fig. 2D~2G).

Cap 58~65 mm, grayish-turquoise or turquoise gray to dark turquoise. Lamellae white, 3~4 mm broad, crowded. Stipe  $120~150 \times 10~12$  mm, cylindrical or slightly tapering upward, white, smooth to slightly scaly, stuffed to slightly hollow. Bulb hollow, 38~50 mm long and 22~38 mm wide. Ring on the upper part of the stem, white, skirt-like, membranous. Basidia 39.1~51.2 × 10.2~12.9 µm, 4-sterigmate, and a simple basal septum. Spores 9.1~11.8 × 5.9~7.5 µm, Q = 1.29~1.77, broadly ellipsoidal to elongate, occasionally oval.

**Specimens examined:** Korea, Gangwon Province, Inje County, Mt. Jeombong, 38°2′28.56″ N, 128°27′58.87″ E, 22 Aug 2014, Young Woon Lim, SFC20140822-29 (GenBank accession Nos. KT779080 for ITS and KT779067 for nLSU); on the ground of mixed woodlands.

**Remarks:** *Amanita girseoturcosa* is characterized by its turquoise cap. This species was transferred from the section *Phalloideae* to the section *Lepidella* [7].

Amanita imazekii T. Oda, C. Tanaka & Tsuda, Mycologia 93: 1231 (2001) (Table 1, Fig. 2H~2J).

Cap 84 mm, grayish creamy. Lamellae white to pale creamy, crowded. Stipe  $190 \times 13 \sim 16$  mm, slightly tapering upward, pruinose to slightly floccose, white. Bulb saccate, white, 54 mm long and 32 mm wide. Ring on the upper part of the stem, white, skirt-like, membranous. Basidia 45.8~64.7  $\times$  9.9~14.1 µm, 4-sterigmate. Clamps present at the bases of basidiole. Spores 9.8~13.2  $\times$  9.2~11.4 µm, Q = 1.01~1.26, globose to broadly ellipsoidal.

**Specimens examined:** Korea, Gangwon Province, Inje County, Mt. Jeombong, 37°58′20.76″ N, 128°25′53.15″ E, 12 Sep 2014, Hae Jin Cho, SFC20140912-30 (GenBank accession Nos. KT779090 for ITS and KT779077 for nLSU); on the ground of mixed woodlands.

**Remarks:** This species is closely related with *A. incarnatifolia*, *A. murrilliana*, and *A. yuaniana* in the nLSU sequence analysis. However, *A. imazekii* has spores that range from being globose to subglobose, while the other species have ellipsoid to elongate spores [24].

*Amanita sepiacea* S. Imai, Bot. Mag., Tokyo 47: 423-432 (1933) (Table 1, Fig. 2K~2M).

Cap 100~150 mm, dark grey to brown to blackish, more dark in the center, dirty white to greyish conical to verrucose warts on cap. Lamellae white and crowded. Stipe

 $150 \sim 215 \times 10 \sim 20$  mm, cylindrical or slightly tapering upward, surface white to dirty white covered with greyish to grey fibrillose squamules. Bulb subglobose to spindle-shaped to turnip-shaped, 25~35 mm wide. Ring on the upper part of the stem, white, skirt-like, membranous. Basidia  $32.3 \sim 40.6 \times 8.8 \sim 11.1 \,\mu$ m, 4-sterigmate, and a simple basal septum. Spores  $6.7 \sim 8.0 \times 4.9 \sim 6.8 \,\mu$ m, Q =  $1.15 \sim 1.49$ , broadly ellipsoidal to ellipsoidal.

**Specimens examined:** Korea, Gangwon Province, Inje County, Mt. Jeombong, 38°2'28.56″ N, 128°27'58.87″ E, 30 Jul 2014, Hae Jin Cho, SFC20140730-17; on the ground of mixed woodlands. 38°2'26.63″ N, 128°28'19.82″ E, 30 Jul 2014, Hae Jin Cho, SFC20140730-25; on the ground of forest with *Quercus* spp. 22 Aug 2014, Young Woon Lim, SFC20140822-49. 15 Sep 2014, Hae Jin Cho, SFC20140912-15 (Genbank accession Nos. KT779086 for ITS and KT779073 for nLSU).

**Remarks:** Amanita sepiacea has a remarkably large fruit body; the cap grew up to 100~150 mm and stipe rose up to 200 mm. This species is morphologically very similar to Amanita excelsa [13] and Amanita fritillaria [28]. However, A. sepiacea was distinguished in our study from A. excelsa and A. fritillaria by ITS and nLSU sequence analyses (Fig. 1) [29, 30].

### ACKNOWLEDGEMENTS

This work was supported by Inje County (Investigation of Inje Biological Resources) and the National Institute of Biological Resources under the Ministry of Environment (Survey and excavate Korean indigenous fungal species, Project No.: NIBR201501205).

## REFERENCES

- 1. Bas C. Morphology and subdivision of *Amanita* and a monograph of its section *Lepidella*. Persoonia 1969;5:285-573.
- Nara K. Ectomycorrhizal networks and seedling establishment during early primary succession. New Phytol 2006;169:169-78.
- 3. Kirk PM, Cannon PF, Minter DW, Stalpers JA. Dictionary of the fungi. 10th ed. Wallingford: CABI Publishing; 2008.
- 4. Jenkins DT, Petersen RH. A neotype specimen for *Amanita muscaria*. Mycologia 1976;68:463-9.
- 5. Weiß M, Yang ZL, Oberwinkler F. Molecular phylogenetic studies in the genus *Amanita*. Can J Bot 1998;76:1170-9.
- 6. Oda T, Tanaka C, Tsuda M. Molecular phylogeny of Japanese *Amanita* species based on nucleotide sequences of the internal transcribed spacer region of nuclear ribosomal DNA. Mycoscience 1999;40:57-64.
- Cai Q, Tulloss RE, Tang LP, Tolgor B, Zhang P, Chen ZH, Yang ZL. Multi-locus phylogeny of lethal *Amanitas*: implications for species diversity and historical biogeography. BMC Evol Biol 2014;14:143.
- 8. Seok SJ, Lim YW, Kim CM, Ka KH, Lee JS, Han SK, Kim SO, Hur JS, Hyun IH, Hong SG, et al. List of mushrooms in Korea. Seoul: The Korean Society of Mycology; 2013.

414 Cho et al.

- 9. Zhang P, Chen ZH, Xiao B, Tolgor B, Bao HY, Yang ZL. Lethal amanitas of East Asia characterized by morphological and molecular data. Fungal Divers 2010;42:119-33.
- Moreno G, Platas G, Peláez F, Bernedo M, Vargas A, Daza A, Santamaría C, Camacho M, de la Osa LR, Manjón JL. Molecular phylogenetic analysis shows that *Amanita ponderosa* and *A. curtipes* are distinct species. Mycol Prog 2008;7:41-7.
- Moncalvo JM, Drehmel D, Vilgalys R. Variation in modes and rates of evolution in nuclear and mitochondrial ribosomal DNA in the mushroom genus *Amanita* (Agaricales, Basidiomycota): phylogenetic implications. Mol Phylogenet Evol 2000;16:48-63.
- 12. Inje-gun Biological Resources. Inje: Inje County; 2011.
- Breitenbach J, Kränzlin F. Fungi of Switzerland. Switzerland. Vol. 4. Lucerne: Mykologia; 1995.
- Rogers SO, Bendich AJ. Extraction of total cellular DNA from plants, algae and fungi. In: Gelvin SB, Schilperoort RA, editors. Plant molecular biology manual. Dordrecht: Springer; 1994. p. 183-90.
- 15. Gardes M, Bruns TD. ITS primers with enhanced specificity for basidiomycetes: application to the identification of mycorrhizae and rusts. Mol Ecol 1993;2:113-8.
- Vilgalys R, Hester M. Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several *Cryptococcus* species. J Bacteriol 1990;172:4238-46.
- Min YJ, Park MS, Fong JJ, Seok SJ, Han SK, Lim YW. Molecular taxonomical re-classification of the genus *Suillus* Micheli ex S.F. Gray in South Korea. Mycobiology 2014;42: 221-8.
- Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S. MEGA5: molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. Mol Biol Evol 2011;28:2731-9.
- Katoh K, Standley DM. MAFFT multiple sequence alignment software version 7: improvements in performance and usability. Mol Biol Evol 2013;30:772-80.

- Stamatakis A. RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. Bioinformatics 2014;30:1312-3.
- Endo N, Gisusi S, Fukuda M, Yamada A. In vitro mycorrhization and acclimatization of Amanita caesareoides and its relatives on Pinus densiflora. Mycorrhiza 2013;23:303-15.
- 22. Yang ZL. Amanita eijii: a new name for Amanita cokeri f. roseotincta. J Jilin Agric Univ 2002;24:32-4.
- 23. Beardslee HC. A new *Amanita* and notes on *Boletus subalbellus*. J Elisha Mitchell Sci Soc 1936;52:105-6.
- Oda T, Tanaka C, Tsuda M. Amanita imazekii: a new species in Amanita section Caesareae. Mycologia 2001;93:1231-4.
- Kim CS, Jo JW, Kwag YN, Kim JH, Shrestha B, Sung GH, Han SK. Taxonomic study of *Amanita* subgenus *Lepidella* and three unrecorded *Amanita* species in Korea. Mycobiology 2013;41:183-90.
- Park MS, Fong JJ, Lee H, Oh SY, Jung PE, Min YJ, Seok SJ, Lim YW. Delimitation of *Russula* subgenus *Amoenula* in Korea using three molecular markers. Mycobiology 2013;41: 191-201.
- Kim CS, Jo JW, Kwag YN, Oh J, Shrestha B, Sung GH, Han SK. Four newly recorded *Amanita* species in Korea: *Amanita* sect. *Amanita* and sect. *Vaginatae*. Mycobiology 2013;41:131-8.
- Sanmee R, Tulloss RE, Lumyong P, Dell B, Lumyong S. Studies on *Amanita* (Basidiomycetes: Amanitaceae) in Northern Thailand. Fungal Divers 2008;32:97-123.
- Zhang L, Yang J, Yang Z. Molecular phylogeny of eastern Asian species of *Amanita* (Agaricales, Basidiomycota): taxonomic and biogeographic implications. Fungal Divers 2004;17:219-38.
- Hosen MI, Li TH, Deng WQ. Amanita cinereovelata, a new species of Amanita section Lepidella from Bangladesh. Mycol Prog 2015;14:35.