

Filamin B Serves as a Molecular Scaffold for Type I Interferon-induced c-Jun NH₂-terminal Kinase Signaling Pathway

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Type I interferons (IFNs) activate Janus tyrosine kinase-signal transducer and activator of transcription pathway for exerting pleiotropic biological effects, including antiviral, antiproliferative, and immunomodulatory responses. Here, we demonstrate that filamin B functions as a scaffold that links between activated Rac1 and a c-Jun NH₂-terminal kinase (JNK) cascade module for mediating type I IFN signaling. Filamin B interacted with Rac1, mitogen-activated protein kinase kinase kinase 1, mitogen-activated protein kinase kinase 4, and JNK. Filamin B markedly enhanced IFN α -dependent Rac1 activation and the sequential activation of the JNK cascade members. Complementation assays using M2 melanoma cells revealed that filamin B, but not filamin A, is required for IFN α -dependent activation of JNK. Furthermore, filamin B promoted IFN α -induced apoptosis, whereas short hairpin RNA-mediated knockdown of filamin B prevented it. These results establish a novel function of filamin B as a molecular scaffold in the JNK signaling pathway for type I IFN-induced apoptosis, thus providing the biological basis for antitumor and antiviral functions of type I IFNs.

INTRODUCTION

Interferons (IFNs) are a family of cytokines that mediate antiviral, antiproliferative, antitumor, and immunomodulatory responses (Boehm *et al.*, 1997; Stark *et al.*, 1998; Kalvakolanu, 2000; Sen, 2000; Taniguchi and Takaoka, 2001; Platanias, 2005). IFNs are divided into two groups: type I (including IFN α and IFN β) and type II (IFN γ). Of these, type I IFNs mediate the pleiotropic biological effects by activating tyrosine kinase (TYK)-2 and Janus tyrosine kinase (JAK)-1 kinases, which are associated with type I interferon receptor (IFNAR) 1 and IFNAR2 subunits of type I IFN receptors, respectively. The activated JAK kinases regulate downstream engagement of multiple signal transducer and activator of transcription (STAT) proteins that translocate to the nucleus to induce gene transcription via binding to the promoters of IFN-stimulated genes.

The first signaling pathway shown to be activated by IFNs is JAK-STAT pathway, but it becomes evident that other IFN-regulated signaling cascades are required for the generation of pleiotropic responses to IFNs. Examples of the

cascades that operate independently of JAK-STAT pathway include mitogen-activated protein kinase (MAPK) signaling pathways that link a variety of extracellular signals to cellular responses as diverse as proliferation, differentiation, and apoptosis (Whitmarsh and Davis, 1998; Garrington and Johnson, 1999; Schaeffer and Weber, 1999; Widmann *et al.*, 1999). Mitogen-activated protein (MAP) kinases are classified into four main groups: p38, extracellular signal-regulated kinase (ERK)1/2, ERK5, and JNK families (Wang and Tournier, 2006). Of MAPK pathways, p38 pathway has most extensively been studied for its role in the generation of IFN-mediated responses. p38 is phosphorylated and activated in a type I IFN-dependent manner and the inhibition of p38 activity blocks IFN α -dependent transcription of genes that are regulated by IFN-stimulated response elements (ISREs) (Goh *et al.*, 1999; Uddin *et al.*, 1999). Moreover, disruption of the *p38 α* gene results in defective transcription of genes that are regulated by ISREs and/or gamma interferon activated sequence elements (Li *et al.*, 2004). The small GTPase Rac1 is activated in a type I IFN-dependent manner, and its function is required for downstream engagement of p38 pathway (Uddin *et al.*, 2000). Type I IFNs also induce the activation of mitogen-activated kinase kinase (MKK) 3 and MKK6 that are essential for p38 activation (Li *et al.*, 2005). Type I IFNs have also been shown to activate JNK for the induction of apoptosis in some lymphoma cells (Yanase *et al.*, 2005), but the downstream signaling cascade that mediates type I IFN-induced JNK activation remains unknown. In addition, it has been reported that ERK2 is activated by type I IFNs and in response to viral infection, and the expression of dominant-negative (dn) ERK2 inhibits type I IFN-induced transcription (David *et al.*, 1995; Wang *et al.*,

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Abbreviations used: ABD, actin-binding domain; GEF, guanine nucleotide-exchange factor; GSH, glutathione; H1, Hinge region 1; IFN, interferon; MAPK, mitogen-activated protein kinase; PARP, poly(ADP-ribose) polymerase; PBD, p21 (GTPase)-binding domain; TRAIL-R1, tumor necrosis factor-related apoptosis-inducing ligand-receptor 1.

2004). However, a recent study has shown that IFN β does not induce ERK activation (Takada *et al.*, 2005).

Filamins are nonmuscle actin-binding proteins that comprise a family of three members: filamin A, B, and C (Stossel *et al.*, 2001; van der Flier and Sonnenberg, 2001). These filamin isoforms are large cytoplasmic proteins that play important parts in cross-linking cortical actin filaments into a dynamic three-dimensional structure. The N-terminal region of filamins contains an actin-binding domain (ABD), followed by a rod-like domain consisting of 24 tandem repeats (Gorlin *et al.*, 1990). Dimerization of filamins through the last C-terminal repeat allows the formation of V-shaped flexible structure that is essential for function (Weihsing, 1988; Gorlin *et al.*, 1990). Filamins interact with >30 cellular proteins of great functional diversity (Stossel *et al.*, 2001). For example, filamin A binds to numerous proteins involved in signal transduction, including the small GTPase RalA, RhoA, Rac1, and Cdc42 (Ohta *et al.*, 1999); the guanine nucleotide exchange factor Trio (Bellanger *et al.*, 2000); and TRAF2 (Leonardi *et al.*, 2000). In addition, the interaction of CD28 with filamin A is required for the induction of T-cell cytoskeletal rearrangements and thus for the recruitment of lipid microdomains and signaling mediators into the immunological synapse (Tavano *et al.*, 2006). Filamin A also serves as an adaptor protein that links human immunodeficiency virus-1 receptors to actin cytoskeleton remodeling machinery for facilitation of virus infection (Jimenez-Baranda *et al.*, 2007). These diverse interactions suggest that filamins function as signaling scaffolds by connecting and coordinating a variety of cellular processes.

In the present study, we demonstrate for the first time that filamin B, but not filamin A, acts as a scaffold that tethers Rac1 and a JNK-specific MAPK module, recruits them to membrane ruffles, and mediates JNK activation in response to type I IFNs. Moreover, filamin B accelerates type I IFN-dependent apoptosis. These results demonstrate that filamin B plays a key role as a molecular scaffold in JNK signaling pathway for type I IFN-induced apoptosis.

MATERIALS AND METHODS

Antibodies and Reagents

Polyclonal anti-filamin A and anti-filamin B antibodies were raised in rabbits by injecting purified proteins. Antibodies against Xpress (Invitrogen, Carlsbad, CA), FLAG, β -actin (Sigma-Aldrich, St. Louis, MO), Rac1 (Millipore, Billerica, MA), hemagglutinin (HA; Roche Diagnostics, Indianapolis, IN), phospho (p)-JNK, JNK, MKK4, p-MKK4, poly(ADP-ribose) polymerase (PARP), c-Myc (Cell Signaling Technology, Danvers, MA), tumor necrosis factor-related apoptosis-inducing ligand-receptor 1 (R&D Systems, Minneapolis, MN), and MEKK1 and glutathione transferase (GST), green fluorescent protein (GFP) (Santa Cruz Biotechnology, Santa Cruz, CA), 6XHis (BD Biosciences, San Jose, CA) were used. Monoclonal anti-Myc antibody was produced from 9E10 hybridoma. Human recombinant IFN α was obtained from R&D Systems.

Plasmid Construction and Mutagenesis

cDNAs for filamin B and its deletion mutants were cloned into pcDNA4-HisMax. Expression vectors for the full-length or the C-terminal hinge-1 to repeat 24 (H1-R24) region of filamin A was also subcloned into pcDNA4-HisMax. Rac1, Rac1/G12V, Rac1/T17N, MEKK1, MEKK4, MKK4, MKK7, JNK1, and p38 cDNAs were subcloned into pcDNA3 containing a 5'-end Myc, FLAG, and/or HA tag sequences. MKK6 and JNK1 cDNAs were subcloned into pcDNA3.1-Myc/His. JNK1, MKK4, MEKK1 Δ , and Rac1 cDNAs were subcloned into pGEX-4T-1. Dominant-negative JNK1 were generated by replacement of Thr183 and Tyr185 by Ala and Phe, respectively.

Sequences for Short Hairpin RNAs (shRNAs)

Two filamin B-specific shRNAs, Rac1-specific shRNA, MEKK1-specific shRNA, MKK4-specific shRNA, and their empty vectors (pSM2c or pLKO.1) were purchased from Open Biosystems (Huntsville, AL). Filamin B-specific shRNAs are tgc tgt tga cag tga gcg acc acc tac ttg gac atc tat ata gtg aag cca cag

atg tat ata gat gtc aaa gta ggt ggg tgc cta ctg cct cgg a (shRNA-1) and tgc tgt tga cag tga gcg ccc agc cag cat cct ttg cta tta gtg aag cca cag atg taa tag cca agg atg ctg gct ggt tgc cta ctg cct cgg a (shRNA-2). Filamin A-specific shRNAs are tgc tgt tga cag tga gcg cgc cca ccc act tca cag taa ata gtg aag cca cag atg tat tta ctg tga agt ggg tgg gct tgc cta ctg cct cgg a (shRNA-1) and tgc tgt tga cag tga gcg ccc acc tac ttg gag atc ttg ata gtg aag cca cag atg tat aaa gat ctc aaa gta ggt ggt tgc cta ctg cct cgg a (shRNA-2). Rac1-specific shRNA is ccg gcg caa aca gat gtg ttc tta act cga gtt aag aac aca tct gtt tgc gtt ttt. MEKK1-specific shRNA is tgc tgt tga cag tga gcg agc ctt tgc tat ctc cat gaa ata gtg aag cca cag atg tat ttc act gag ata cga aag gcc tgc cta ctg cct cgg a. MKK4-specific shRNA is ccg gct tct tat gga ttg gga tgc act cga gta cat cca aat cca taa gaa gtt ttt. MKK6-specific shRNA is ccg ggg cct aca tac cca gag cta act cga gtt agc tct ggg tat gta ggc ctt ttt. Cdc42-specific shRNA is ccg gcc ctc tac tat tga gaa act tct cga gaa gtt tct caa tag tag agg gtt ttg g.

Cell Culture and Transfections

HeLa and A549 cells were cultured in DMEM (JBI, Daegu, Korea) supplemented with 10% fetal bovine serum (FBS). M2 cells that were kindly provided by Dr. T. P. Stossel were grown in minimal essential medium supplemented with 8% normal calf serum and 2% FBS (Cunningham *et al.*, 1992). Cells were transfected using Lipofectamine reagent in combination with PLUS reagent (Invitrogen). All transfection experiments were performed at least three times, and the efficiency of transfection was at least 65% in each experiment. For knockdown of proteins, HeLa cells were transfected with shControl or shRNAs by using jetPEI (Polyplus-transfection, New York, NY). After transfection, cells were treated with puromycin (3 μ g/ml) for 1 wk to select transfected cells. Efficiency of protein knockdown was in range of 70–100% as verified by immunoblot and densitometric analysis.

Immunoprecipitation and Pull-Down Analysis

For immunoprecipitation, cells were lysed in 50 mM Tris-HCl, pH 8.0, containing 150 mM NaCl, 1% Triton X-100, or 0.5% NP-40, 1 mM phenylmethylsulfonyl fluoride (PMSF), and 1 \times protease inhibitor cocktail (Roche Diagnostics). Cell lysates were incubated with appropriate antibodies for 2 h at 4 $^{\circ}$ C and then with 50 μ l of 50% slurry of protein A-Sepharose for 1 h. For pull-down analysis, cell lysates were prepared as described above and treated with nickel-nitrilotriacetic acid (Ni-NTA) agarose (QIAGEN) and glutathione (GSH)-Sepharose (GE Healthcare, Little Chalfont, Buckinghamshire, United Kingdom).

In Vitro Binding Assay

GST-tagged Rac1, MEKK1 Δ , MKK4, JNK, and GST were expressed in *Escherichia coli*, Rosetta (DE3) strain, and purified using GSH-Sepharose. The C-terminal R20-24 region of filamin B was subcloned into pET32a, expressed in Rosetta strain, and purified using NTA agarose. For NTA pull-down analysis, 3 μ g of His-R20-24 was incubated 3 μ g of GST or GST-tagged Rac1, MEKK1 Δ , MKK4, or JNK in buffer A consisting of 50 mM Tris-HCl, pH 8.0, 150 mM NaCl, 0.5% NP-40, 10 mM Imidazole, 1 mM PMSF, and 1 \times protease inhibitor cocktail for 1 h at 4 $^{\circ}$ C and then with 30 μ l of 50% slurry of NTA resins for the next 1 h. Precipitated beads were washed three times with buffer A containing 20 mM imidazole. They were then subjected to SDS-polyacrylamide gel electrophoresis (PAGE) followed by immunoblot with anti-GST antibody. For GST-pull-down analysis, the GST- and His-tagged proteins were incubated as described above but in buffer B consisting of 50 mM Tris-HCl, pH 8.0, 150 mM NaCl, 0.5% NP-40, 1 mM PMSF, and 1 \times protease inhibitor cocktail for 1 h at 4 $^{\circ}$ C and then with 30 μ l of 50% slurry of GSH-Sepharose for the next 1 h. Precipitated beads were washed three times with buffer B containing 250 mM NaCl. They were then subjected to SDS-PAGE by immunoblot with anti-6XHis antibody.

In Vitro Kinase Assay

For assaying JNK activity, cell lysates were incubated for 5 h with GST-cJun bound to GSH-Sepharose beads. Precipitates were washed twice with buffer C consisting of 20 mM Tris-HCl, pH 7.4, 150 mM NaCl, 1 mM EDTA, 1 mM EGTA, 1% Triton X-100, 2.5 mM sodium pyrophosphate, 1 mM β -glycerophosphate, 1 mM Na $_3$ VO $_4$, 1 μ g/ml leupeptin, and 1 mM PMSF. They were again washed with buffer D consisting of 25 mM Tris-HCl, pH 7.5, 5 mM β -glycerophosphate, 2 mM DTT, 0.1 mM Na $_3$ VO $_4$, and 10 mM MgCl $_2$. After washing, precipitates were incubated with 0.2 mM ATP in buffer D for 30 min at 30 $^{\circ}$ C. The samples were resolved by SDS-PAGE, and phosphoproteins were visualized by immunoblot with anti-p-cJun antibody.

For assaying MEKK1 activity, cell lysates were subjected to immunoprecipitation with anti-MEKK1 antibody. Precipitates were incubated with 2 μ g of GST-MKK4 as a substrate in buffer D containing 0.2 mM ATP for 30 min at 30 $^{\circ}$ C. The samples were then resolved by SDS-PAGE, and phosphoproteins were visualized by immunoblot with anti-p-MKK4 antibody.

Rac1 Activation Assay

Rac1 activation by IFN α was assayed by measuring its ability to interact with GST-fused GTPase-binding domain of Pak1 (GST-PBD) (Benard *et al.*, 1999). Briefly, the GTPase-binding domain of human Pak1 was expressed in *E. coli*

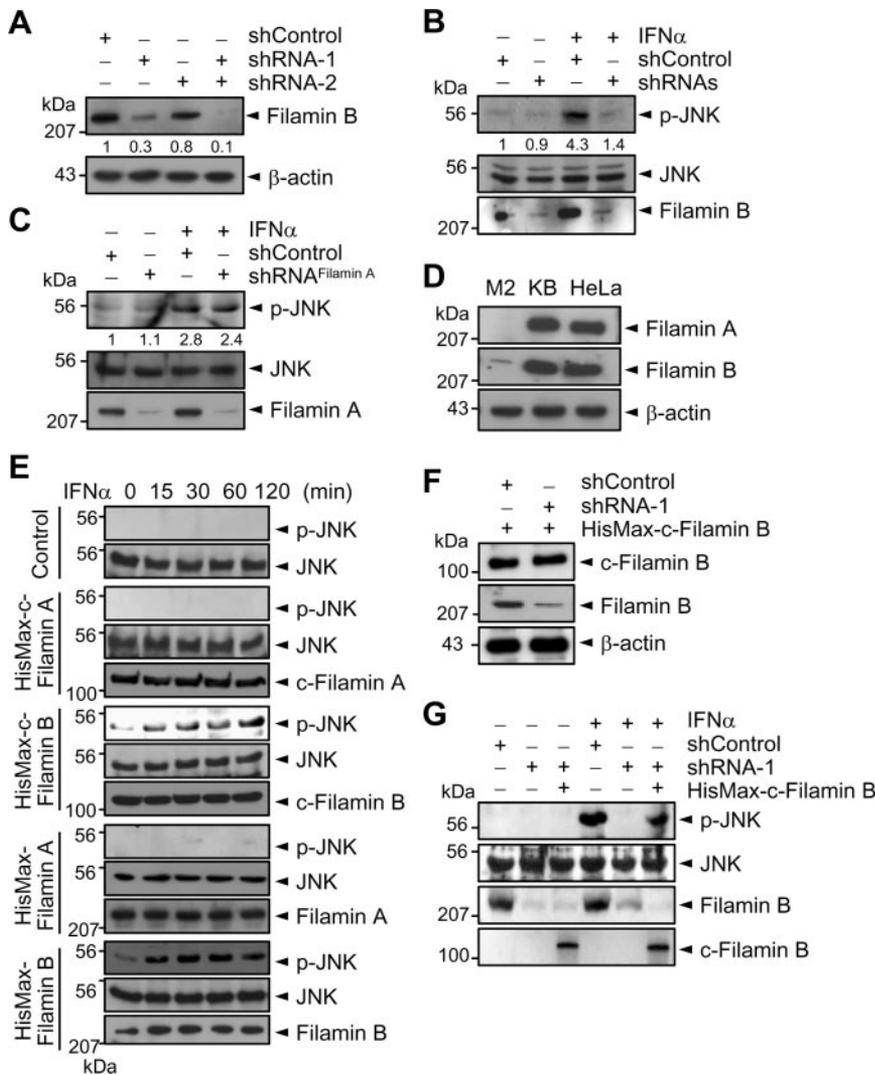


Figure 1. Filamin B is required for type I IFN-induced JNK activation. (A) HeLa cells were transfected with either or both of filamin B-specific shRNA-1 and -2 or with shControl. Cell lysates were subjected to immunoblot with anti-filamin B or anti- β -actin antibody. (B) HeLa cells transfected with shRNAs or shControl were cultured for 1 h with or without IFN α . Cell lysates were subjected to immunoblot with anti-p-JNK or anti-JNK antibody. (C) Experiments were performed as in B but using filamin A-specific shRNAs or shControl. In A–C, the numerals indicate the densitometrically quantified band intensities relative to that seen with shControl only and are representative of three independent experiments. (D) Lysates from the indicated cells were subjected to immunoblot with anti-filamin A or anti-filamin B antibody. (E) M2 cells were transfected with a vector expressing HisMax-c-filamin B, HisMax-c-filamin A, HisMax-tagged full-length filamin A or filamin B, or an empty vector (Control). They were then cultured with IFN α . (F) HisMax-c-filamin B was expressed in HeLa cells transfected with shRNA-1 or shControl. Cell lysates were subjected to immunoblot with anti-filamin B or anti-Xpress antibody. (G) HeLa cells prepared as in F were cultured for 1 h with or without IFN α . Cell lysates were subjected to immunoblot with anti-p-JNK or anti-JNK antibody. The data in D–F are representative of three independent experiments, which showed similar results.

as a GST fusion protein. Cells were serum-starved for 3 h, treated with 10,000 U/ml IFN α for the next 0.5 h, and lysed in buffer C. Cell lysates were incubated for 1 h with 5 μ g of GST-PBD-conjugated Sepharose beads. Bound proteins were pulled down, separated by SDS-PAGE, and subjected to immunoblot with anti-Rac1 antibody.

Immunocytochemistry

Cells were grown on coverslips. Two days after transfection, they were fixed by incubation for 10 min with 3.7% paraformaldehyde in phosphate-buffered saline (PBS). Cells were washed three times with PBS containing 0.1% Triton X-100, permeabilized with 0.5% Triton X-100 in PBS for 5 min, and treated with 3% bovine serum albumin (BSA) in PBS for 1 h. They were then incubated for 1 h with appropriate antibodies. After washing with PBS containing 0.1% Triton X-100, cells were incubated for 1 h with fluorescein isothiocyanate-, tetramethylrhodamine B isothiocyanate-, or Cy5-conjugated secondary antibody in PBS containing 3% BSA. To visualize F-actin, fixed cells were also stained with rhodamine-phalloidin. Cells were then observed using a confocal laser scanning microscope (LSM510; Carl Zeiss, Jena, Germany). Images were processed using Photoshop (Adobe Systems, Mountain View, CA).

RESULTS

Filamin B Is Required for Type I IFN-induced JNK Activation

To determine whether filamin B is involved in type I IFN-induced activation of JNK, HeLa cells were transfected with filamin B-specific shRNA-1 (directed against N-terminal re-

peat 2), shRNA-2 (against C-terminal repeat 22), or both. Immunoblot analysis reveals that each shRNA, but not a negative control (shControl), could reduce the protein level of endogenous filamin B (Figure 1A). The filamin B level was further reduced upon transfection of both shRNA-1 and -2 (henceforth referred to as shRNAs). Furthermore, knock-down of endogenous filamin B by shRNAs markedly reduced IFN α -induced activation of JNK (Figure 1B). Because filamin A shows high sequence similarity with filamin B, we examined whether filamin A-specific shRNAs might also prevent IFN α -induced JNK activation. However, knock-down of endogenous filamin A showed little or no effect on JNK activation (Figure 1C). These results implicate a crucial role of filamin B, but not filamin A, in type I IFN-induced JNK signaling.

M2 is a human malignant melanoma cell line that does not express filamin A (Cunningham *et al.*, 1992). Moreover, filamin B level in M2 cells was <5% of that in HeLa or KB cells (Figure 1D). To clarify further the involvement of filamin B in type I IFN-induced JNK activation, the C-terminal region from H1-R24 (see Figure 6A for the structural organization of filamin B) was expressed in M2 cells. Henceforth, the C-terminal H1-R24 region of filamin B was referred to as c-filamin B unless otherwise indicated. Likewise, the C-

subjected to assay for Rac1 activation. The level of activated Rac1 was increased by IFN α treatment, and this Rac1 activation was further enhanced by c-filamin B (Figure 2E). Furthermore, knockdown of filamin B by shRNAs led to a marked inhibition of IFN α -induced Rac1 activation (Figure 2F), implicating the role of filamin B in IFN α -induced Rac1 activation. These results also suggest that filamin B serves as a scaffold that recruits Rac1 for mediating type I IFN signaling.

Filamin B Colocalizes with Rac1 in Membrane Ruffles

Because filamin B promotes type I IFN-induced Rac1 activation and because activated Rac1 is known to participate in membrane ruffle formation (Allen *et al.*, 1997), we examined whether filamin B could be recruited to the ruffles in response to IFN α . Without IFN α , both endogenous filamin B and Rac1 showed a diffused cytoplasmic localization in HeLa cells (Figure 3A). In IFN α treated cells, a significant portion of them with F-actin was redistributed to and colocalized in the cell periphery, where membrane ruffles formed. These results suggest that type I IFNs promote not only the interaction between Rac1 and filamin B but also their colocalization in membrane ruffles. We then examined whether the knockdown of filamin B influences membrane ruffle formation. Transfection of filamin B-specific shRNA-1 led to a marked reduction in Rac1-mediated ruffle formation in IFN α -treated cells, whereas it showed little or no effect on the cytoplasmic distribution of Rac1 in untreated cells (Figure 3B). Figure 3C confirms the knockdown of filamin B by shRNA-1 in the cells used for the immunocytochemical analysis. In addition, the fractions of cells, showing colocalization of filamin B with Rac1 in membrane ruffles, were quantified (Figure 3D). These results suggest that filamin B promotes Rac1-mediated membrane ruffle formation in response to type I IFNs.

To demonstrate that the interaction of Rac1 with filamin B is required for the promotion of Rac1-mediated ruffle formation in response to type I IFNs, we generated an in-frame deletion of R20-21 in HisMax-c-filamin B for preventing its interaction with Rac1 (termed c- Δ R20-21; Figure 6A). HisMax-c-filamin B and HisMax-c- Δ R20-21 were expressed in HeLa cells that had been depleted of endogenous filamin B by shRNA-1. Reconstitution of c-filamin B, but not c- Δ R20-21, led to a marked increase in the formation of membrane ruffles, where c-filamin B and Rac1 colocalized with enriched F-actin in cells treated with IFN α (Figure 4). These results are consistent with our findings that HisMax-c-filamin B is capable of activating Rac1 in response to IFN α (Figure 2E). Thus, it seems clear that filamin B plays an important role in the promotion of Rac1-mediated ruffle formation through its interaction with and activation of Rac1. Because HisMax-c-filamin B lacks ABD, these results also suggest that ABD, which is known to play a role in actin organization, is not required for the colocalization of filamin B with Rac1 in membrane ruffles.

Filamin B Interacts with JNK Cascade Members

To determine whether filamin B interacts with a JNK-specific signaling module, HA- or Myc-tagged MAPK cascade members were expressed in HeLa cells with or without HisMax-c-filamin B. HisMax-c-filamin B was coimmunoprecipitated with MEKK1, MKK4, and JNK1 (Figure 5A). However, HisMax-c-filamin B was not coprecipitated with MEKK4 or MKK6 that is a p38-specific activator (data not shown). HisMax-c-filamin B was neither coprecipitated with MKK7 that also is a JNK activator (data not shown), suggesting that filamin B shows a binding selectivity even among MAP2Ks that activate JNK. In addition, all of Rac1,

MEKK1, MKK4, and JNK1 could coprecipitate HisMax-tagged full-length filamin B (Figure 5B). Collectively, these results suggest that filamin B serves as a scaffold, tethering a JNK-specific MAPK module that is downstream to Rac1 (Rac1 \rightarrow MEKK1 \rightarrow MKK4 \rightarrow JNK1). Marti *et al.* (1997) have reported that filamin A interacts with MKK4 and does not directly bind MEKK1 or JNK, and thereby suggested that MKK4 may form a ternary complex with filamin A and MEKK1. They also showed that filamin A-deficient M2 cells are defective in tumor necrosis factor- α -stimulated JNK activation and this defect could be recovered by complementation of filamin A. Therefore, it would be of interest to compare the ability of filamin A with that of filamin B in their interaction with JNK cascade members. HisMax-c-filamin A or HisMax-c-filamin B was expressed in HeLa cells with each of Myc-tagged MEKK1, MKK4, and JNK1 or Myc-Rac1. Immunoprecipitation analysis reveals that both c-filamin A and c-filamin B could interact with Rac1, MEKK1, and MKK4 (Figure 5C). However, c-filamin B showed a much higher affinity to MEKK1 than c-filamin A. Furthermore, c-filamin A, unlike c-filamin B, could not interact with JNK1. These results are consistent with our finding that filamin A could not promote type I IFN-induced JNK activation.

To determine whether endogenous filamin B could interact with endogenous JNK cascade members and type I IFNs affect this interaction, HeLa cells were incubated with or without IFN α . Immunoprecipitation analysis reveals that filamin B could be coprecipitated with MEKK1, MKK4, and JNK (Figure 5D) and vice versa (Figure 5E), but only when cells were incubated with IFN α . These results suggest that endogenous filamin B plays a role as a scaffold tethering JNK cascade members as well as Rac1 (Figure 2C) in response to type I IFNs. We then examined whether filamin B could directly bind JNK cascade members. His-R20-24 and GST-tagged MEKK1 Δ , MKK4, and JNK1 were expressed in *E. coli* and purified to apparent homogeneity. NTA pull-down analysis shows that all of the JNK cascade members could be coprecipitated with His-R20-24 (Figure 5F). Reciprocally, pull-down analysis using GSH-Sepharose beads reveals that His-R20-24 could be coprecipitated with the JNK cascade members. Together, these results indicate that MEKK1, MKK4, and JNK1 physically interact with filamin B.

To map the regions within filamin B for the interaction with Rac1 and JNK cascade members, various deletions of filamin B were generated (Figure 6A) and expressed in HeLa cells with Myc-tagged Rac1, MEKK1, MKK4, or JNK1. HisMax-tagged R20-24 and H1-R21, but not R22-24, could be coimmunoprecipitated with Rac1 (Figure 6B). Moreover, HisMax-c- Δ R20-21 having in-frame deletion of R20-21 in HisMax-c-filamin B could not interact with Rac1, indicating that R20-21 is essential for the binding of Rac1. Alternatively, HisMax-tagged R18-24 and R20-24, but not H1-R21, could be coprecipitated with MEKK1 (Figure 6C), suggesting that the binding site of MEKK1 lies within R20-24. Both MKK4 and JNK1 interacted with R18-24, R20-24, and H1-R21. These results suggest that the binding site for MKK4 and JNK1 to filamin B resides within R20-21.

Filamin B Serves as a Scaffold for JNK Cascade Members

To determine whether filamin B indeed acts as a scaffold that tethers JNK cascade members for type I IFN-induced JNK activation, M2 cells were complemented with HisMax-c-filamin B and then treated with IFN α . NTA pull-down analysis shows that IFN α treatment leads to a dramatic increase in the interaction of c-filamin B with all of endogenous Rac1, MEKK1, MKK4, and JNK (Figure 7A). Moreover, immuno-

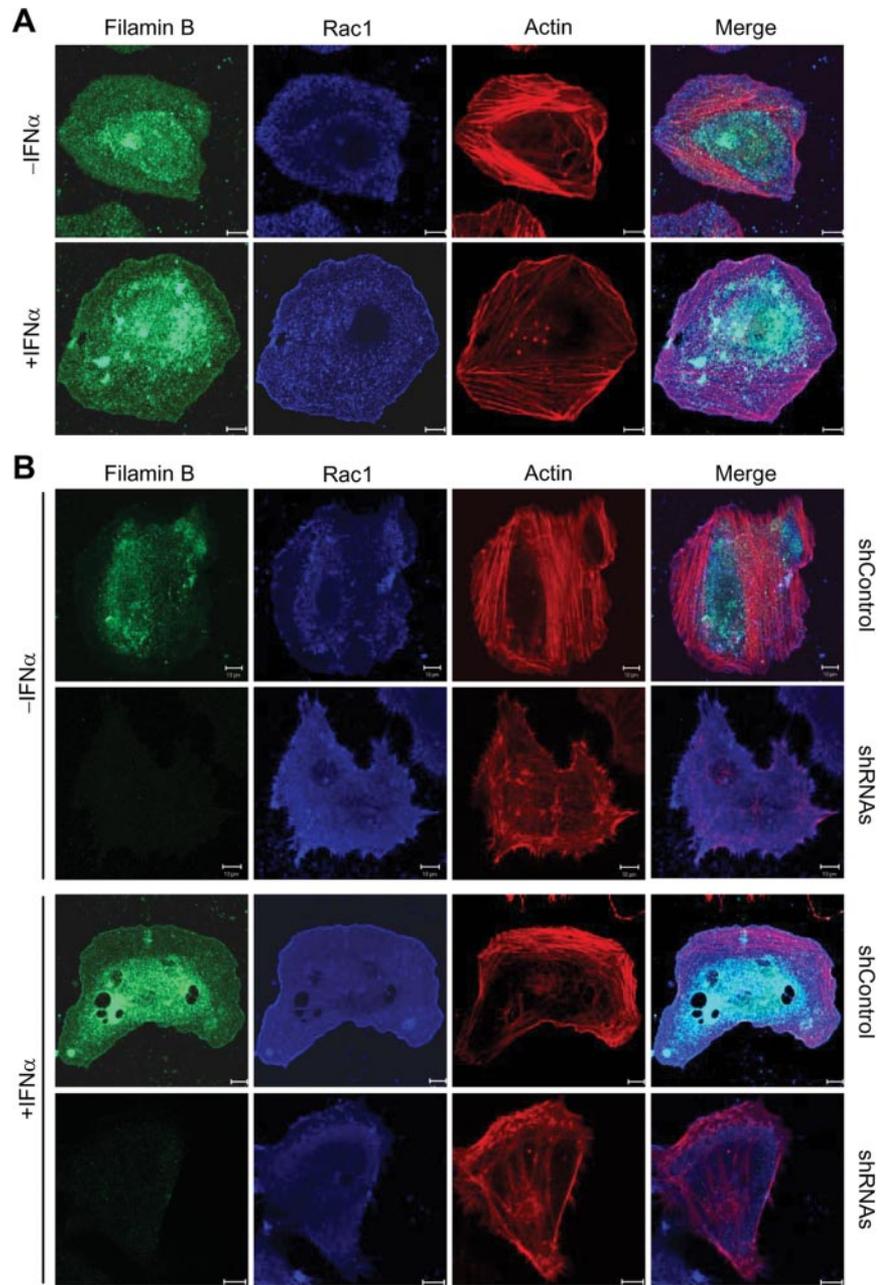
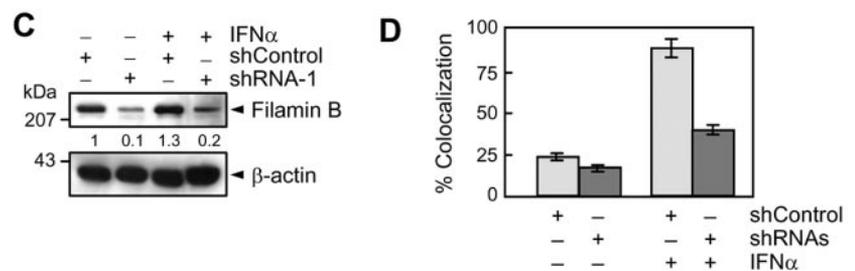


Figure 3. Filamin B colocalizes with Rac1 in membrane ruffles. (A) HeLa cells incubated with or without IFN α for 30 min were stained with anti-filamin B or anti-Rac1 antibody or phalloidin. The bars indicate 10 μ m. (B) Cells transfected with shRNA-1 or shControl were cultured and stained as in A. Bars, 10 μ m. (C) Lysates were obtained from the same cells used in B and subjected to immunoblot with anti-filamin B or anti- β -actin antibody. The numerals indicate the quantified band intensities relative to that seen with shControl only and are representative of three independent experiments. (D) The fractions of cells showing colocalization of filamin B with Rac1 in membrane ruffles were quantified. Error bars are \pm SEM of total 90 cells from at least three independent experiments.



blot analysis of the same precipitates and lysates reveals that the level of p-JNK increases in parallel with the increase in the interaction of c-filamin B with JNK cascade members. Together with the data shown in Figure 5E, these results indicate that filamin B serves as a scaffold that tethers JNK cascade members and that type I IFNs promote the forma-

tion of filamin B scaffold complex. To clarify further whether filamin B can form a complex with endogenous JNK cascade members, HisMax-c-filamin B was expressed in M2 cells. After incubation with IFN α for 1 h, cell lysates were subjected to gel filtration. Immunoblot analysis reveals that all of the endogenous Rac1, MEKK1, MKK4, and JNK proteins

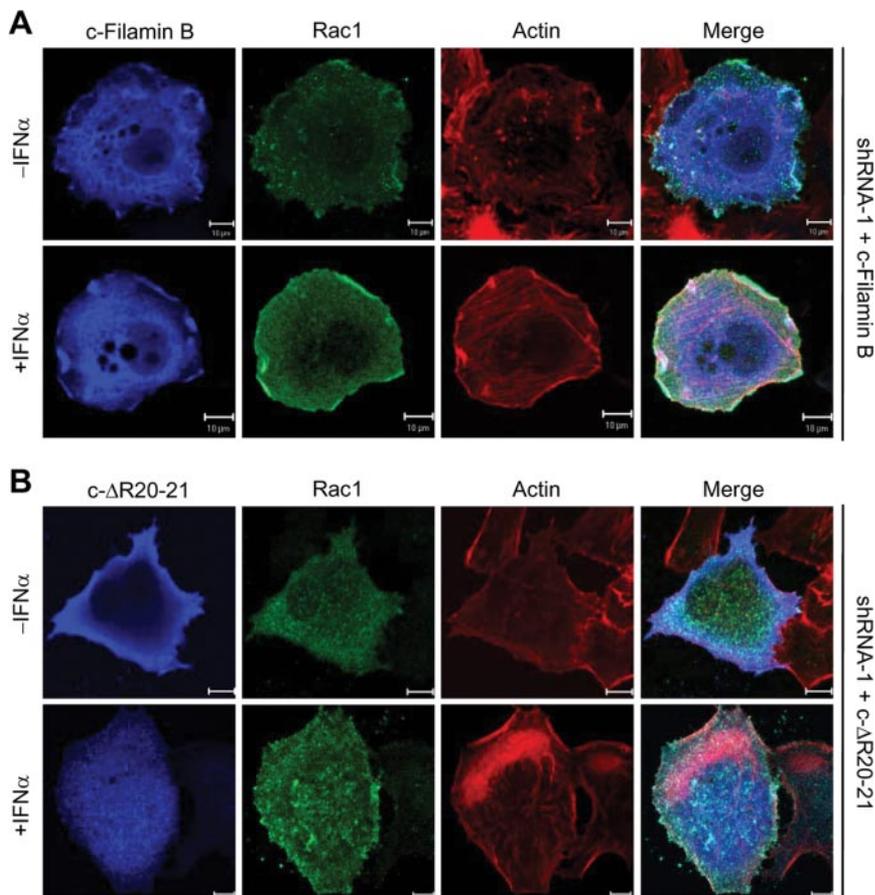


Figure 4. Requirement of the interaction between filamin B and Rac1 for their colocalization in membrane ruffles. HisMax-c-filamin B (A) or HisMax-c- Δ R20-21 (B) was expressed in HeLa cells that had been transfected with shRNA-1. Cells were then incubated with or without IFN α for 30 min followed by staining with anti-Xpress or anti-Rac1 antibody or phalloidin. Bars, 10 μ m.

cofractionate with c-filamin B when M2 cells were complemented with HisMax-c-filamin B, but not without it (Figure 7B). These results confirm that filamin B serves as a scaffold for Rac1 and the JNK cascade members.

Of note was the finding that all of the proteins obtained from M2 cells without complementation of HisMax-c-filamin B ran on the column with sizes even larger than their original sizes. Particularly, MEKK1 behaved as a much larger protein than that from M2 cells complemented with HisMax-c-filamin B. MEKK1 is known to associate with vesicles and thus shows up as speckles (Fanger *et al.*, 1997). Therefore, we examined whether the decrease in the apparent size of MEKK1 by filamin B complementation might be due to a change in its subcellular localization (i.e., dissociation from the vesicles and subsequent binding to filamin B in membrane ruffles). In accord with the previous report, MEKK1 showed up as speckles when expressed alone (Figure 7C). However, upon coexpression with HisMax-c-filamin B, a significant portion of MEKK1 was recruited to membrane ruffles where c-filamin B concentrated. Moreover, MKK4 and JNK1, which by themselves locate throughout the cytoplasm and the nucleus, were also recruited to membrane ruffles when coexpressed with HisMax-c-filamin B. Similarly, Rac1 that alone resides in the cytoplasm and cell periphery were concentrated in the ruffles upon HisMax-c-filamin B coexpression. Thus, the alteration in MEKK1 behavior on the gel filtration column seems to be due to the change in its subcellular localization upon c-filamin B complementation. Likewise, the changes in the chromatographic behavior of Rac1, MKK4, and JNK could be due to the alterations in their interaction partners under the same conditions. These results also implicate a critical role of filamin B as a scaffold that sequesters Rac1

and JNK cascade members in membrane ruffles for facilitating the type I IFN signaling.

Filamin B Promotes Sequential Activation of JNK Cascade Members

To determine the effect of filamin B on sequential activation of JNK cascade members, Myc-MKK4 was expressed in HeLa cells with or without HA-MEKK1. Immunoprecipitation analysis revealed that expression of MEKK1 promoted the phosphorylation of MKK4, and this MEKK1 activity was markedly enhanced by coexpression of HisMax-c-filamin B (Figure 8A). To determine the effect of filamin B on MEKK1- and MKK4-mediated activation of JNK1, JNK1-Myc-His was expressed in cells with HA-MEKK1 or Myc-MKK4. The level of p-cJun was increased by MEKK1 expression, and this increase in JNK1 activity was again enhanced by coexpression of HisMax-c-filamin B (Figure 8B). Likewise, expression of both MKK4 and HisMax-c-filamin B activated JNK1 to a greater extent than that of MKK4 alone (Figure 8C). These results indicate that filamin B is capable of promoting the MEKK1- and MKK4-mediated JNK1 activation by serving as a scaffold of the effector kinases.

Deletion analysis of filamin B revealed that the binding site of MEKK1 lies within R20-24 and that of JNK1 and MKK4 resides in R20-21 (Figure 6C). To confirm the role of filamin B as a scaffold in JNK signaling pathway, in-frame deletions of full-length filamin B were generated in the regions that have the binding sites for MEKK1 and JNK1 (Figure 6A). We then examined their ability to interact with MEKK1 and JNK1 by expressing them in HeLa cells followed by immunoprecipitation analysis. Myc-JNK1 was coprecipitated with filamin B that lacks either R20 or R21

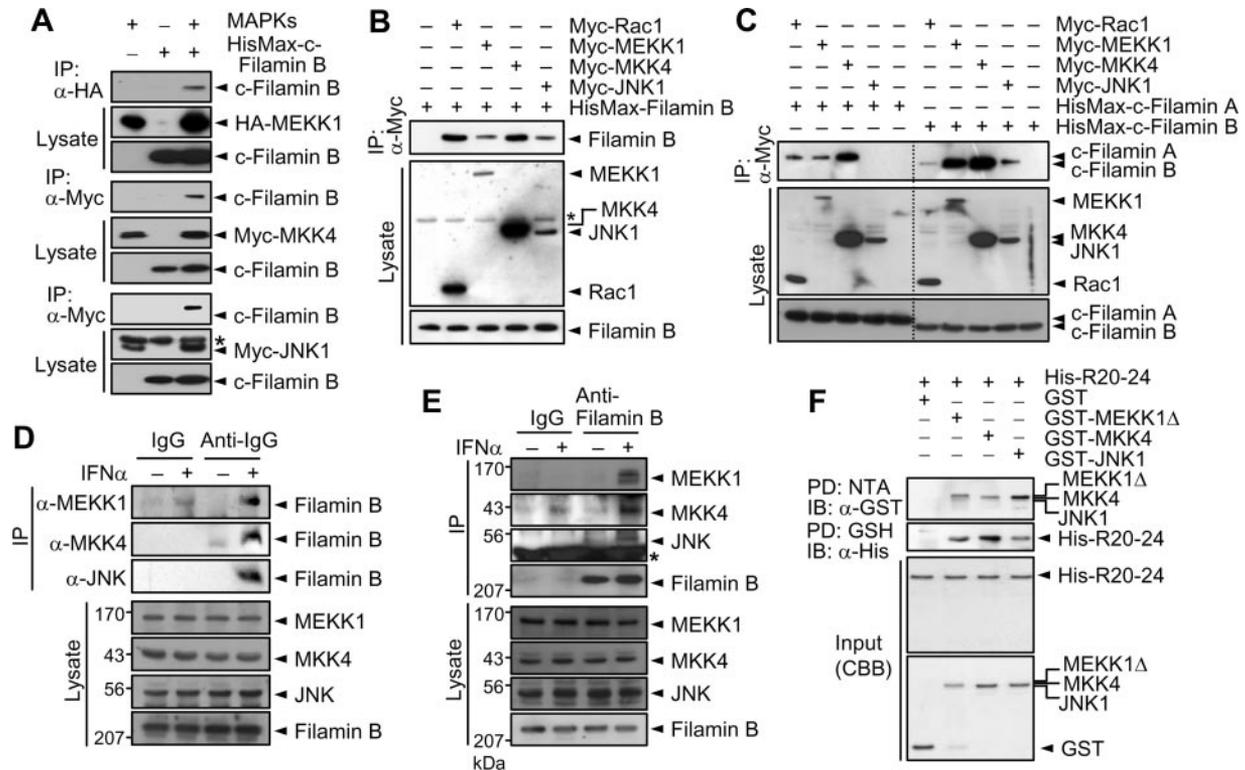


Figure 5. Filamin B interacts with JNK cascade members. (A) HA-MEKK1, Myc-MKK4, or Myc-JNK1 was expressed in HeLa cells with HisMax-c-filamin B. Cell lysates were subjected to immunoprecipitation with anti-HA or anti-Myc antibody followed by immunoblot with anti-Xpress antibody. The asterisk in the bottom panel indicates IgG heavy chain. (B) Myc-tagged Rac1, MEKK1, MKK4, or JNK1 was expressed in HeLa cells with HisMax-tagged full-length filamin B. Cell lysates were subjected to immunoprecipitation with anti-Myc antibody followed by immunoblot with anti-Xpress antibody. The asterisk indicates a nonspecific band. (C) Myc-tagged Rac1, MEKK1, MKK4, or JNK1 was expressed in HeLa cells with HisMax-c-filamin A or HisMax-c-filamin B. Cell lysates were subjected to immunoprecipitation as described in B. (D) HeLa cells were incubated with or without IFN α for 1 h. Cell lysates were then subjected to immunoprecipitation with anti-MEKK1, anti-MKK4, or anti-JNK antibody or normal rabbit IgG (IgG) followed by immunoblot with anti-filamin B antibody. (E) Cells were incubated as in D, and their lysates were subjected to immunoprecipitation with anti-filamin B or preimmune serum (IgG) followed by immunoblot with anti-MEKK1, anti-MKK4 or anti-JNK antibody. (F) GST-tagged MEKK1 Δ , MKK4, and JNK1 was incubated with His-R20-24 for 1 h at 4°C. His-R20-24 was pulled down with NTA resins, and precipitates were subjected to SDS-PAGE followed by immunoblot with anti-GST antibody (top). From the same incubation mixtures, GST-tagged proteins were pulled down with GSH-Sepharose, and precipitates were subjected to immunoblot with anti-His antibody (second panel). Purified GST-tagged proteins and His-R20-24 were subjected to SDS-PAGE followed by staining with CBB. Note that we used a deletion mutant of MEKK1 (MEKK1 Δ) having the amino acid sequence of 397-750 for pull-down analysis, because its full-length form could not be expressed in *E. coli*. Similar data were obtained by at least three independent trials of each experiment set.

(HisMax- Δ R20 or HisMax- Δ R21, respectively), but not with that lacking both R20 and R21 (HisMax- Δ R20-21), indicating that JNK1 can bind either R20 or R21 (Figure 8D). Myc-MEKK1 was also coprecipitated with HisMax- Δ R20, but not with HisMax- Δ R21 or HisMax- Δ R20-21, indicating that R21, but not R20, is essential for the binding of MEKK1 to filamin B (Figure 8E). Consistently, in-frame deletion of R21 or both R20 and R21 led to a marked decrease in the ability of filamin B to promote IFN α -induced JNK activation (Figure 8F). Furthermore, overexpression of HisMax-R20-24 in HeLa cells dramatically inhibited the ability of endogenous filamin B to promote IFN α -induced JNK activation (Figure 8G), most likely by sequestering Rac1 and JNK cascade members into separate complexes. Collectively, these results confirm that filamin B plays an essential role as a scaffold in type I IFN-induced JNK signaling pathway.

Filamin B Is Required for IFN α -induced Activation of JNK Cascade Members

To determine whether endogenous filamin B indeed plays an essential role in the activation of JNK cascade members in

response to type I IFNs, HeLa cells were transfected with shRNAs or shControl. IFN α caused a dramatic activation of endogenous Rac1 (Figure 2F), MEKK1 (Figure 9A), MKK4 (Figure 9B), and JNK (Figure 1B), and these activations were abrogated by transfection of filamin B-specific shRNAs. To determine whether filamin B serves as a link between activated Rac1 and JNK cascade for mediating type I IFN signaling, Rac1/G12V was expressed in cells transfected with shControl or shRNAs. Knockdown of filamin B led to a marked reduction in cJun phosphorylation (Figure 9C). These results indicate that filamin B serves as a scaffold that recruits endogenous Rac1 and JNK cascade members for efficient and facilitated activation of JNK in response to IFN α .

To determine whether the upstream effectors of JNK are indeed required for type I IFN-induced activation of JNK, HeLa cells transfected with shRNA specific to Rac1, MEKK1, or MKK4 were cultured with IFN α . IFN α markedly induced JNK activation in shControl-transfected cells, whereas little or no JNK activation could be seen in cells transfected with shRNA specific to Rac1, MEKK1, or MKK4 (Figure 9, D–F). As controls, cells were also transfected with MKK6- or

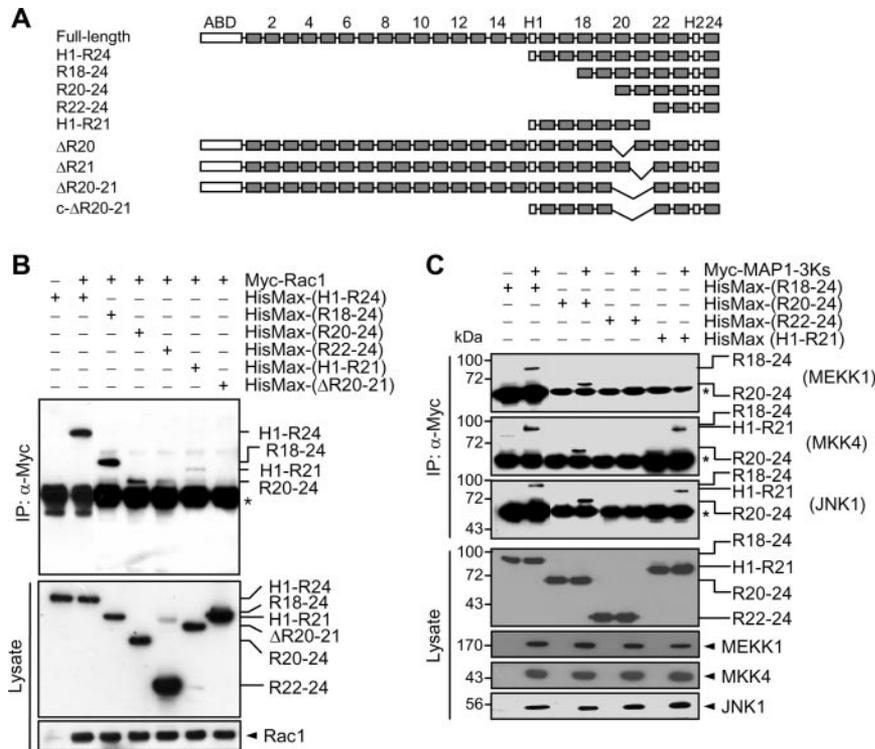


Figure 6. Mapping of the binding regions for Rac1 and JNK cascade members within filamin B. (A) Serial and in-frame deletion mutants of filamin B were generated and subcloned into pcDNA4-HisMax vector. ABD denotes the actin-binding domain, H1 and H2 indicate the hinge regions, and the numerals show the repeat numbers. (B and C) HisMax-tagged deletions of filamin B were expressed in HeLa cells with Myc-Rac1 (B) or with Myc-tagged-MEKK1, MKK4, or JNK1 (C). Cell lysates were subjected to immunoprecipitation with anti-Myc antibody followed by immunoblot with anti-Xpress antibody. The asterisks indicate IgG heavy chain. Nearly the same data were obtained by two independent trials of each experiment.

Cdc42-specific shRNA. However, knockdown of MKK6 or Cdc42 did not show any effect on IFN α -induced JNK activation (Figure 9G). These results indicate that Rac1, MEKK1, and MKK4 are essential components of type I IFN-induced JNK signaling pathway.

Filamin B Accelerates Type I IFN-induced Apoptosis

Type I IFNs induce apoptosis in part through up-regulation of TRAIL and its receptor TRAIL-R1/R2 (Oshima *et al.*, 2001), and this up-regulation depends on IFN-induced JNK activation (Yanase *et al.*, 2005). To determine whether filamin B promotes type I IFN-induced apoptosis, HeLa cells transfected with shRNAs or shControl were treated with IFN α . IFN α treatment led to an increase in the levels of TRAIL-R1 as well as in the cleavage of PARP in cells transfected with shControl (Figure 10A). However, knockdown of filamin B by shRNAs abrogated both IFN α -induced up-regulation of TRAIL-R1 and PARP cleavage, indicating that filamin B accelerates apoptosis. To determine whether filamin B-promoted apoptosis is indeed mediated by IFN α -induced JNK activation, HisMax-c-filamin B was expressed in M2 cells with or without dominant-negative form of JNK1 (dn-JNK1). Expression of dn-JNK1 strongly attenuated PARP cleavage (Figure 10B), indicating that filamin B accelerates IFN α -induced apoptosis through JNK activation. Together, these results demonstrate that filamin B accelerates type I IFN-induced apoptosis by promoting JNK activation.

DISCUSSION

In the present study, we have demonstrated that filamin B serves as a scaffold that tethers Rac1 and a JNK-specific MAPK module (Rac1 \rightarrow MEKK1 \rightarrow MKK4 \rightarrow JNK) and mediates JNK activation in response to type I IFNs. Moreover, filamin B was shown to mediate type I IFN-induced apoptosis through the activation of JNK signaling pathway.

These findings establish a novel function of filamin B as a scaffold in JNK signaling pathway for type I IFN-induced apoptosis. In contrast to filamin B, filamin A is known to mediate cell survival under stress conditions, such as stretch-induced physical stress (Glogauer *et al.*, 1998) and genotoxic stresses (Yuan and Shen, 2001). In addition, filamin A has been shown to integrate with β 1 integrins to mediate cell spreading and is required for adhesion-dependent cell survival (Kim *et al.*, 2008). Although the molecular basis for the opposite biological effects of filamin A and filamin B remains unclear, a notable difference is that under both in vivo and in vitro conditions filamin A, unlike filamin B, is unable to bind JNK that plays a pivotal role in apoptosis. In addition, filamin A, unlike filamin B, does not physically bind MEKK1 (Marti *et al.*, 1997), although both filamin isoforms can interact with MEKK1 under in vivo conditions albeit with marked different affinities (Figure 5C), suggesting that in vivo interaction between filamin A and MEKK1 is indirect. Thus, it is likely that filamin isoforms possess distinct functional specificity in serving as scaffolds by tethering different sets of context-specific signaling molecules in response to different stimuli.

An intriguing, but unanswered, question is on the mechanism by which Rac1, MEKK1, MKK4, and JNK1 interact with filamin B in an IFN α -dependent manner under in vivo conditions, despite our finding that they can directly bind to filamin B under in vitro conditions. Müller *et al.* (2001) have shown that KSR1 translocates from the cytoplasm to the cell surface in response to growth factors and this process is regulated by C-TAK1. In unstimulated cells, C-TAK1 associates with KSR1 and phosphorylates Ser392 to confer 14-3-3 binding and cytoplasmic sequestration of the scaffold protein. On stimulation, the phosphorylation state of Ser392 is reduced, allowing the KSR1 complex to colocalize with activated Ras and Raf-1 at the plasma membrane, thereby facilitating the activation of mitogen-activated protein ki-

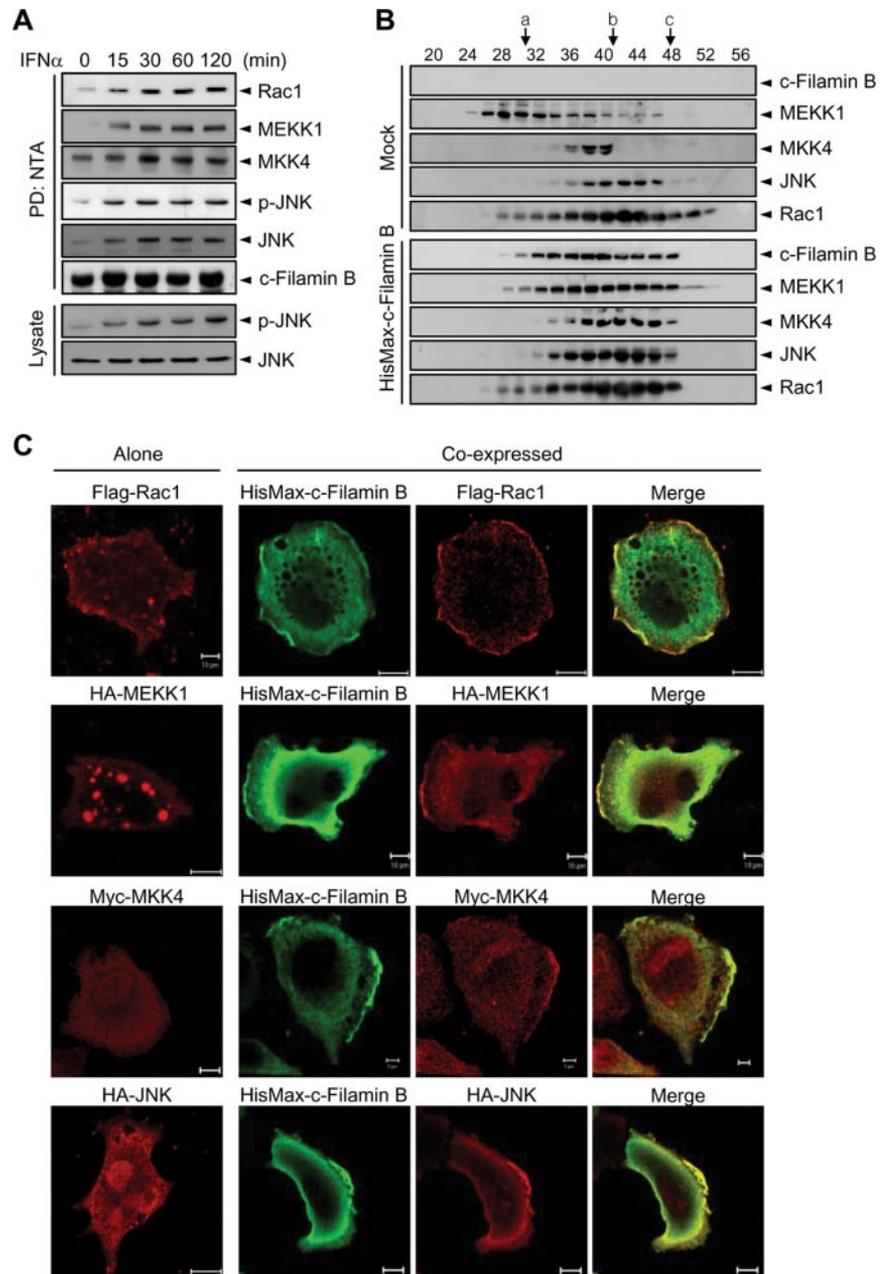


Figure 7. Filamin B serves as a scaffold for JNK cascade members. (A) M2 cells complemented with HisMax-c-filamin B were treated with IFN α . Cell lysates were subjected to pull-down with NTA resins followed by immunoblot analysis. The same precipitates and lysates were also probed with anti-p-JNK or anti-JNK antibody. (B) M2 cells transfected with an empty or HisMax-c-filamin B vector were cultured for 1 h with IFN α . Cell lysates were subjected to gel filtration on a Sephacryl S-300 column (0.7 \times 28 cm) equilibrated with 20 mM Tris-HCl, pH 8.0, containing 150 mM NaCl and 7% glycerol. Fractions of 0.2 ml were collected and subjected to immunoblot with anti-Xpress antibody or antibody against Rac1, MEKK1, MKK4, or JNK. The size markers used were a, thyroglobulin ($M_r = 669,000$); b, β -amylase (200,000); and c, albumin (66,000). (C) FLAG-Rac1, HA-MEKK1, Myc-MKK4, or HA-JNK1 was expressed in HeLa cells with or without HisMax-c-filamin B. Cells were fixed and stained with anti-FLAG, anti-Xpress, anti-HA, or anti-Myc antibody. Bars, 10 μ m. Similar data were obtained by at least three independent trials of each experiment set.

nase kinase and MAPK. Therefore, it is tempting to speculate that modification of filamin B, such as phosphorylation/dephosphorylation, in response to type I IFNs may be involved in the control of its ability to interact with Rac1 and JNK cascade members.

The existence of multiple JNK-specific MAP2Ks and MAP3Ks within a single cell raises an issue regarding the specificity of JNK activation. It has been suggested that the specificity of JNK activation may be achieved by forming signaling complexes on a specific scaffold protein. Indeed, a variety of scaffold proteins for JNK signaling pathway have been identified (Weston and Davis, 2002; Morrison and Davis, 2003; Dard and Peter, 2006). Examples include β -arrestin-2, CrkII, IKAP, JIP1-4, MKPX, POSH, and SKRP1. JIP1 and JIP2 are closely related proteins that bind to JNK, MKK7, and MLKs. Thus, it seems that in addition to MAPK cascade modules, their upstream effectors, such as small

GTPases and guanine nucleotide-exchange factors (GEFs), play a critical role in conferring the specificity of JNK activation in response to specific signals.

The Rho family of small GTPases plays essential roles in the control of a variety of important cellular processes, including actin cytoskeleton rearrangement, cell cycle progression, and cellular transformation (Hall, 1998; Jaffe and Hall, 2002). Moreover, evidence has emerged that small GTPases are often organized into functional modules by scaffold proteins that interact with and tether key components of each signaling pathway. For examples, OSM scaffold, through its ability to bind the GTPase Rac, the actin filaments, and the upstream kinases MEKK3 and MKK3, activates p38 in response to osmestress (Uhlík *et al.*, 2003). POSH also seems to function as a scaffold that links activated Rac1 and JNK cascade to promote neuronal cell death (Xu *et al.*, 2003). In this study, we show that type I IFNs dramatically increase the interaction

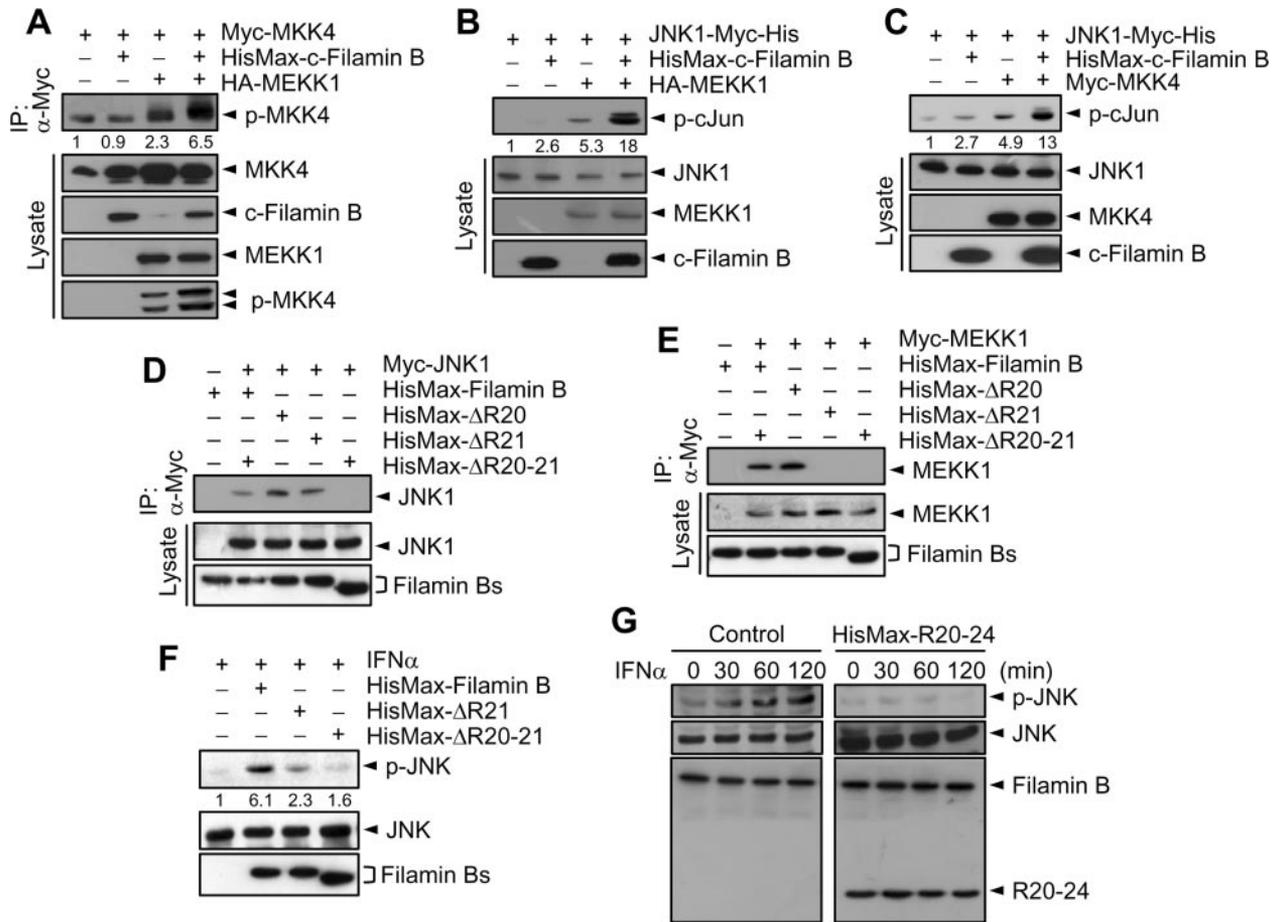


Figure 8. Filamin B promotes sequential activation of JNK cascade members. (A) Myc-MKK4 was expressed in HeLa cells with HisMax-c-filamin B, HA-MEKK1, or both. Cells lysates were subjected to immunoprecipitation with anti-Myc antibody followed by immunoblot with respective antibodies. (B) JNK1-Myc-His was expressed in HeLa cells with HA-MEKK1, HisMax-c-filamin B, or both. Cell lysates were assayed for the phosphorylation of cJun, followed by immunoblot with anti-p-cJun antibody. (C) JNK1-Myc-His was expressed in HeLa cells with Myc-MKK4, HisMax-c-filamin B, or both. (D and E) HisMax-tagged full-length filamin B or its mutant form having in-frame deletion of R20 (Δ R20), R21 (Δ R21), or both R20 and R21 (Δ R20-21) was expressed in HeLa cells with Myc-JNK1 (D) or Myc-MEKK1 (E). Cell lysates were then subjected to immunoprecipitation with anti-Myc antibody. (F) HisMax-tagged filamin B, Δ R21, or Δ R20-21 was complemented to M2 cells. After incubation with IFN α for 1 h, cell lysates were subjected to immunoblot with anti-p-JNK, anti-JNK, or anti-Xpress antibody. (G) HeLa cells were transfected with an empty (control) or HisMax-R20-24 vector. After incubating them with IFN α for various periods, cell lysates were subjected to immunoblot with anti-p-JNK, anti-JNK, anti-filamin B, or anti-Xpress antibody. In A–C, the numerals indicate the quantified band intensities relative to that seen with MKK4 or JNK alone. In F, the numerals indicate the quantified band intensities relative to that seen with transfection of an empty vector. All the numerals are representative of three independent experiments.

between endogenous filamin B and Rac1 and that this interaction leads to a marked enhancement of Rac1 activation. Furthermore, filamin B was shown to play a critical role as a scaffold that links between activated Rac1 and JNK cascade to promote apoptosis in response to type I IFNs. Thus, it seems that upon binding to filamin B, Rac1 serves as an upstream effector that mediates type I IFN signals to JNK signaling module for JNK activation.

Of note was the finding that Rac1 interacts with filamin B independently of its nucleotide-bound state. Several scaffold proteins are known to interact with specific GEFs for the activation of JNK pathway. The scaffold protein hCNK1 links Rho and RhoGEFs to JNK activation (Jaffe *et al.*, 2005). Although JIPs do not seem to bind Rac1 or Cdc42, RacGEFs Tiam1 and Ras-GRF1 bind to JIP2 for selective stimulation of p38 upon Rac activation (Buchsbbaum *et al.*, 2002). These GEFs also bind to JIP1, which may lead to enhanced JNK activation in different cellular context (Morrison and Davis, 2003). Therefore, we propose that filamin B also binds a spe-

cific GEF that controls the activation state of Rac1 for transducing type I IFN signals from IFN receptors to JNK-specific cascade. That is, filamin B may provide a platform for GEF to turn on Rac1 so that GTP-bound Rac1 can initiate type I IFN-dependent JNK signaling pathway. In this regard, it would not be necessary for filamin B itself to distinguish between GDP- and GTP-bound forms of Rac1 for binding. In addition, MEKK1 was shown to regulate JNK pathway in a Cdc42/Rac-dependent manner (Fanger *et al.*, 1997). Thus, MEKK1 may associate with GTP-bound Rac1 on filamin B scaffold for subsequent activation of downstream kinases.

Another possibility for the control of the activation state of Rac1 could be the involvement of upstream protein kinases that regulate the GEF activity. It has been reported that type I IFN-induced activation of Rac1 in p38 pathway could be regulated by upstream tyrosine kinases, such as JAKs or other IFN α -dependent kinases downstream to JAKs, through phosphorylation and activation of GEF for Rac1 (Mayer *et al.*, 2001; Uddin *et al.*, 2000). Among GEFs, Vav is

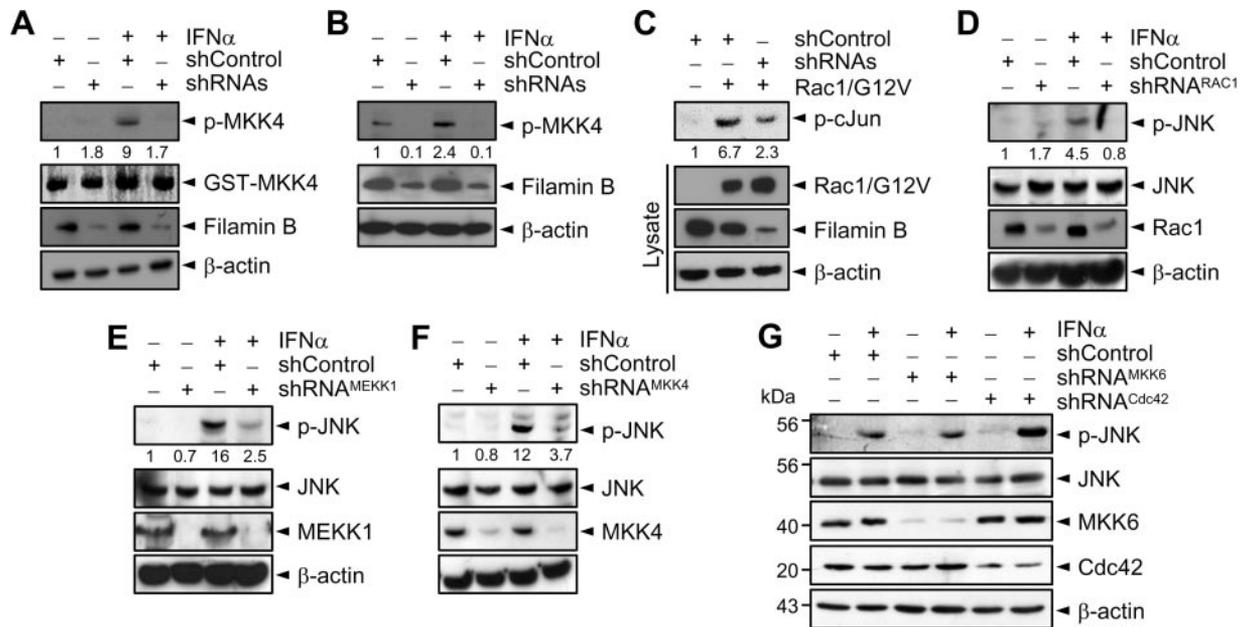


Figure 9. Filamin B is required for IFN α -induced activation of JNK cascade. (A) HeLa cells transfected with shRNAs or shControl were incubated for 1 h with or without IFN α . Cell lysates were immunoprecipitated with anti-MEKK1 antibody. Precipitates were then subjected to in vitro kinase assay for MEKK1 by using GST-MKK4 as a substrate. (B) Cells were incubated as in A, and their lysates were immunoblotted with anti-p-MKK4. (C) A vector expressing Rac1/G12V was transfected to HeLa cells that had been transfected with shRNAs or shControl. Cell lysates were assayed for cJun phosphorylation. (D–F) HeLa cells transfected with shRNA specific for Rac1 (D), MEKK1 (E), or MKK4 (F) were incubated for 1 h with or without IFN α . Cell lysates were then subjected to immunoblot with anti-p-JNK or anti-JNK antibody. In A–F, the numerals indicate the quantified band intensities relative to that seen with shControl only and are representative of three independent experiments. (G) HeLa cells transfected with shRNA specific for MKK6 or Cdc42 were incubated for 1 h with or without IFN α . Cell lysates were then subjected to immunoblot with respective antibodies.

tyrosine-phosphorylated in an IFN α -dependent manner (Micouin *et al.*, 2000; Plataniias and Sweet, 1994; Uddin *et al.*, 1997). Furthermore, Vav has been shown to function as a GEF for Rac1 and via Rac1 activation, regulate the p38 and

JNK signaling pathways (Crespo *et al.*, 1997; Gringhuis *et al.*, 1998; Salojin *et al.*, 1999). Thus, Vav could be a potential candidate that regulates the activation state of Rac1 for transducing type I IFN-induced JNK signaling.

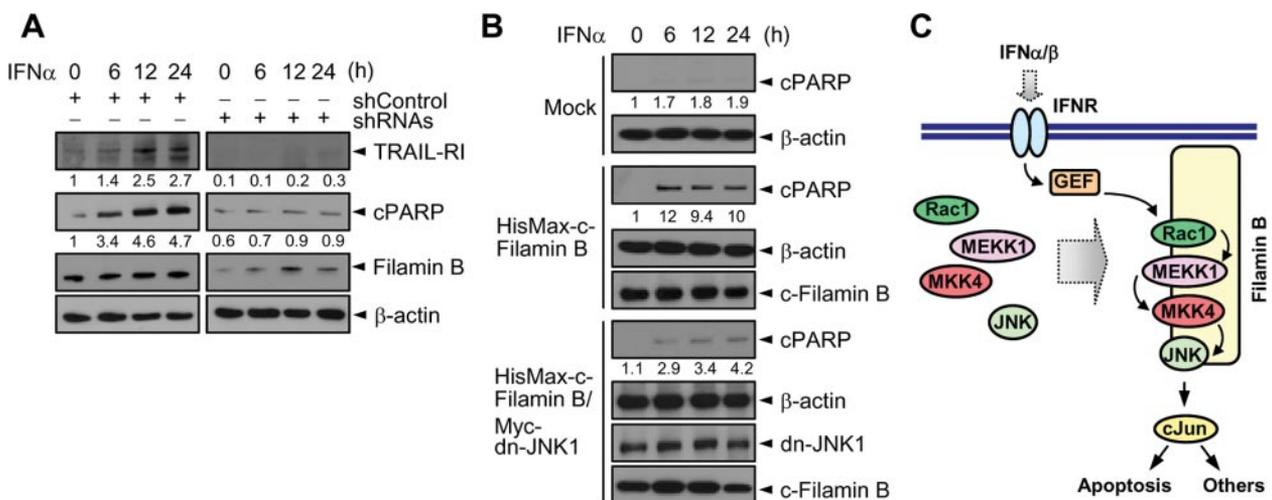


Figure 10. Filamin B accelerates type I IFN-induced apoptosis. (A) HeLa cells transfected with shRNAs or shControl were treated with IFN α . Cell lysates were immunoblotted with anti-TRAIL-R1, anti-PARP, anti-Xpress, or anti- β actin antibody. (B) HisMax-c-filamin B was expressed in M2 cells with or without Myc-dn-JNK1. After incubation with IFN α , cell lysates were subjected to immunoblot analysis. Mock indicates cells transfected with empty vectors. In A and B, the numerals indicate the ratio of quantified intensity of each band to that of β -actin, relative to that seen at the zero time of IFN α treatment, and they are representative of three independent experiments. (C) Binding of type I IFNs to their receptor (IFNR) induces the recruitment of filamin B to membrane ruffles and the binding of Rac1, MEKK1, MKK4, and JNK1 to filamin B scaffold. GEF, a putative guanine nucleotide exchange factor, may then specifically link IFN signal to Rac1 for the successive activation of downstream kinases and thereby for the generation of type I IFN-mediated biological responses, such as apoptosis.

An important characteristic of scaffolds is their unique subcellular localization with specific signaling modules, thus allowing the facilitated interaction between signaling components and the efficient regulation of respective signaling pathway. Remarkably, expression of HisMax-c-filamin B led to the recruitment of Rac1, MEKK1, MKK4, and JNK1 to membrane ruffles (Figure 7C). Furthermore, IFN α treatment resulted in colocalization of endogenous filamin B and Rac1 in membrane ruffles and reconstitution of HisMax-c-filamin B to filamin B-depleted cells restored ruffle formation. These results suggest that filamin B plays an important role in the sequestration of Rac1 and JNK cascade members to membrane ruffles, where they form a complex with filamin B scaffold for efficient transduction of type I IFN signaling.

However, this finding raises a question how HisMax-c-filamin B (i.e., H1-R24) lacking ABD can promote ruffle formation that is known to require dynamic actin rearrangements. It has been established that Pak1 is activated by GTP-bound Rac1 and activated Pak1 plays a key in ruffle formation (Edwards *et al.*, 1999). Filamin A is a substrate of Pak1 and required for Pak1-mediated ruffle formation. In retrospect, filamin A stimulates the Pak1 activity in response to growth factors, indicating that Pak1 and filamin A mutually influence the dynamic actin cytoskeletal structure (Vadlamudi *et al.*, 2002). Furthermore, the Pak1-binding site resides in the C-terminal region of filamin A and a filamin A fragment lacking ABD can promote ruffle formation (Dyson *et al.*, 2001; He *et al.*, 2003). Similarly, HisMax-c-filamin B was found to interact with Pak1 (data not shown). Thus, it seems that H1-R24 stimulates ruffle formation by promoting type I IFN-induced Rac1 activation, which in turn activates Pak1, although it is unknown whether filamin B is phosphorylated by Pak1.

An additional question that needs to be answered is how filamin B, an actin-binding protein, carries out its signaling function without ABD. However, previous studies have shown that the actin-binding property mediated by ABD of filamin A is dispensable in numerous cases for its signaling function. For examples, complementation of N-terminally truncated filamin A in M2 cells could rescue transforming growth factor- β responsiveness (Sasaki *et al.*, 2001), normal trafficking of large-conductance Ca²⁺-activated K⁺ channels to the plasma membrane (Kim *et al.*, 2007), and μ opioid receptor down-regulation (Onoprishvili *et al.*, 2008). Furthermore, C-terminal filamin A fragments could naturally be generated by proteolytic cleavage between R15 to R16, and the cleavage site is conserved in other filamin isoforms (Ozanne *et al.*, 2000; Loy *et al.*, 2003). In addition, generation of filamin A fragments could be regulated by a phosphorylation/dephosphorylation process (Jay *et al.*, 2004; Garcia *et al.*, 2006). Therefore, it has been proposed that proteolysis of scaffold proteins could provide a general mechanism to integrate cellular pathway. Together, it seems possible that type I IFNs induce the phosphorylation and cleavage of filamin B and the resulting C-terminal fragments (e.g., H1-R24) may participate in IFN-signaling pathway.

Until present, the only scaffold protein known to participate in type I IFN signaling is RACK1, which promotes the JAK-STAT pathway by forming a multiprotein complex consisting of the IFN α receptor, STAT1, JAK1, and TYK2 (Usacheva *et al.*, 2003). Thus, filamin B represents the first scaffold that was shown in this study to mediate type I IFN-induced MAPK signaling. Type I IFNs are cytokines that exert important biological activities, including antiviral, antiproliferative, and immunomodulatory responses. These properties have led to the use of type I IFNs in the clinical treatment of certain malignancies, such as chronic myelogenous leukemia, hairy cell leukemia, lymphomas, and certain solid tu-

mors (Platanias, 2003). The potent antiviral properties of IFNs have also ignited clinical studies, which have established their usefulness such as in the viral hepatitis syndromes in humans (Liang *et al.*, 2000; Melian and Plosker, 2001; Regev and Schiff, 2001). Despite the clinical uses of IFNs and the substantial advances in the field of IFN signaling, the precise mechanism by which IFNs mediate their biological effects and relative contribution of each of the various IFN-activated signaling pathways in the induction of specific-IFN responses remain unclear. Therefore, our finding of filamin B that serves as a scaffold for accelerating type I IFN-induced apoptosis should contribute to the understanding of molecular mechanisms by which IFNs mediate the antitumor or antiviral activities of IFNs.

In summary, we propose a model for the role of filamin B in type I IFN-induced signaling pathway that leads to JNK activation as shown in Figure 10C. Binding of IFN α/β to type I IFN receptor activates an unknown GEF, resulting in the activation of Rac1. Activated Rac1 then plays a role in the activation of MEKK1 possibly through their interaction on filamin B scaffold, and thus for successive phosphorylation and activation of MKK4 and JNK. Activated JNK subsequently phosphorylates multiple downstream effectors, like cJun, for the generation of type I IFN-dependent biological responses, such as apoptosis. Together, our data establish that filamin B as a scaffold plays an essential role in type I IFN signaling pathway by mediating the formation of Rac1-MEKK1-MKK4-JNK phospho-relay module.

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